A Rational Approach to Perioperative Fluid Management

Daniel Chappell, M.D.,* Matthias Jacob, M.D.,* Klaus Hofmann-Kiefer, M.D.,* Peter Conzen, M.D.,† Markus Rehm, M.D.‡

Replacement of assumed preoperative deficits, in addition to generous substitution of an unsubstantiated increased insensible perspiration and third space loss, plays an important role in current perioperative fluid regimens. The consequence is a positive fluid balance and weight gain of up to 10 kg, which may be related to severe complications. Because the intravascular blood volume remains unchanged and insensible perspiration is negligible, the fluid must accumulate inside the body. This concept brings into question common liberal infusion regimens. Blood volume after fasting is normal, and a fluid-consuming third space has never been reliably shown. Crystalloids physiologically load the interstitial space, whereas colloidal volume loading deteriorates a vital part of the vascular barrier. The endothelial glycocalyx plays a key role and is destroyed not only by ischemia and surgery, but also by acute hypervolemia. Therefore, undifferentiated fluid handling may increase the shift toward the interstitial space. Using the right kind of fluid in appropriate amounts at the right time might improve patient outcome.

PERIOPERATIVE fluid application has been a topic of debate in past years. After the ongoing controversy on colloids versus crystalloids¹⁻³ and proposing the ideal composition of saline fluids,⁴⁻⁷ the main focus is now on the amount of applied fluids in general.⁸⁻¹³ The discussion is still dominated by the advocates of a more liberal regimen.¹⁴⁻²⁰ Most perioperative fluid overload is regarded as a minor problem, and studies showing increased fluid accumulation in tissue have not changed this attitude.^{21,22} Rather, preoperative volume loading is considered indispensable by many,^{15,19,23-26} and fluid boluses are part of most recommendations for perioperative care.^{11,27} This statement is mainly based on four generally unquestioned pathophysiologic "fundamentals": (1) The preoperatively fasted patient is hypovolemic because of ongoing insensible perspiration and urinary output¹⁰; (2) the insensible perspiration increases dramatically when the surgeon starts cutting the skin barrier²⁷; (3) an unpredictable fluid shift toward the third space requires generous substitution²⁸; and (4) hypervolemia is harmless because the kidneys regulate the overload.29

The purpose of this review is to promote a rational perioperative fluid management, combining common knowledge with clinical research results and new phys-

Mark A. Warner, M.D., served as Handing Editor for this article.

iologic insights regarding the vascular barrier. This review will explain and underline the importance, quantity, and destination of perioperative fluid shifting and its related problems.

Perioperative Fluid Optimization: Do All Roads Lead to Rome?

The attending anesthesiologist is faced daily with several principal and practical problems when arranging perioperative fluid handling. Under normal circumstances, the individual patient's hydration and volume state before surgery is unknown. In addition, the exact target remains unclear, and many theoretically possible targets cannot be measured in clinical routine. The principal goal is to optimize cardiac preload. An important determinant, total body blood volume, should be optimized to achieve this. Importantly, optimizing does not necessarily mean *maximizing*, despite frequently being interpreted in this way,³⁰ and blood volume cannot be assessed routinely: Double-label blood volume measurement, the current standard to assess total body blood volume, is invasive, complex, and personnel intensive.³¹⁻³⁴ Alternative methods that do not use sampling lack calibration and are, therefore, imprecise.³⁵ Hematocrit dilution is often based on estimated basic values and can only assess changes in the circulating part of the blood volume,^{33,36,37} ignoring a considerable noncirculating portion of the plasma (see section titled The Endothelial Glycocalyx: The Gateway to the Interstitial Space). Therefore, direct blood volume measurements are possible in principle and are frequently used to answer scientific questions. Unfortunately, they remain impractical in everyday routine.

Measuring volume responsiveness, occasionally referred to as a "goal-directed" approach, 30, 38, 39 seems at first to be an interesting alternative to directly measuring blood volume, but it has several limitations. First, there is no proof that this circulatory surrogate, enabling the clinician to maximize stroke volume, really achieves the optimum. Second, the two still most applied measures in this context, *i.e.*, pulmonary capillary wedge pressure and central venous pressure, do not at all predict volume responsiveness, in clear contrast to the common assumption.^{40,41} Systolic pressure and pulse pressure variation, on the contrary, predict volume responsiveness, but do not improve patient outcome.38 Stroke volume maximization via esophageal Doppler-guided fluid boluses seems to improve outcome,³⁰ especially in elderly and frail patients.³⁹ This method, however, cannot be performed everywhere and in every patient for practical and

 $^{^{\}star}$ Staff Anesthesiologist, † Professor of Anesthesiology, ‡ Associate Professor of Anesthesiology.

Received from the Clinic of Anesthesiology, Ludwig-Maximilians University, Munich, Germany. Submitted for publication August 6, 2007. Accepted for publication April 8, 2008. Support was provided solely from institutional and/or departmental sources. Drs. Chappell and Jacob contributed equally to this work.

Address correspondence to Dr. Jacob: Clinic of Anesthesiology, Ludwig-Maximilians University Munich, Nussbaumstrasse 20, 80336 Munich, Germany. matthias.jacob@med.uni-muenchen.de. This article may be accessed for personal use at no charge through the Journal Web site, www.anesthesiology.org.

financial reasons. Performing esophageal Doppler measurements in awake patients, to apply a rational fluid regimen from the very beginning of the anesthesiologic procedure, is almost impossible. Moreover, it has up until now been compared with only standard fluid handling, which revealed no large differences between the total fluid volume of the two studied groups.^{30,42} Accordingly, assuming the worst case, the actual message behind these data could also be that esophageal Dopplerguided fluid overload is superior to uncontrolled fluid overload. But this message does not answer current questions regarding alternative concepts.

"Liberal," "Standard," or "Restrictive": It's in the Eye of the Beholder

Results of studies on fluid therapy will have an impact on everyday practice only if clinicians are able to accept one or more alternative regimens as being superior. Many clinicians are reluctant to change their fluid practices, impeding research on perioperative fluid handling and acceptance of protocol-based improvements. Research suffers not only from an almost unascertainable target, but traditionally from a lack of standardization, complicating the design of control and study groups. Investigators have normally named their traditional regimen the standard group and compared it with their own restrictive ideas. Consequently, a restrictive regimen in one study is often designated as liberal in another setup. In addition, studies claiming to compare restrictive versus liberal use of fluid should, in part, rather be interpreted as investigating hypovolemia versus normovolemia.¹³ This shortcoming prevents even promising results from impacting daily clinical routine and makes any pooling of the data impossible. A further important limitation of the data in this field is the target of a given study. Perioperative fluid handling has been related to, among other things, nausea and vomiting, pain, tissue oxygenation, cardiopulmonary disorders, need of revision surgery, duration of hospital stay, and bowel recovery time. However, the relevance of each individual target depends on the examined type and extent of surgery, which in turn has an enormous influence on changes and significance of these outcome parameters. Avoiding postoperative nausea and vomiting (PONV) in cardiopulmonary-healthy patients, for example, might be the most important goal after a 15-min knee arthroscopy. By contrast, it is merely a minor issue after a 6-h major abdominal intervention, in which cardiopulmonary complications or mortality rates are in the spotlight. Therefore, a careful differentiation between large and small operations, as well as abdominal versus nonabdominal surgery, seems to be necessary.

Major Surgery

Even though the results of several studies regarding major nonabdominal surgery are currently underpowered and partly inconclusive,⁴³ the findings in patients receiving major abdominal surgery are quite promising. It has been demonstrated that protocol-based fluid restriction reduced the incidence of perioperative complications such as cardiopulmonary events^{8,11} and disturbances of bowel motility,11,44 while improving wound and anastomotic healing^{8,11} and reducing hospital stay.^{11,44} Lobo et al.⁴⁴ investigated 20 adults after elective colonic resection. Intraoperative fluid application was quite aggressive (20 ml \cdot kg⁻¹ \cdot h⁻¹) in all patients, but postoperatively, they were randomly assigned to either a restrictive (≤ 21 per day) or a standard (≥ 31 per day) protocol. The latter caused a significant weight gain, a later return of bowel function, and a prolonged hospital stay. It seems as though not only intraoperative but also postoperative fluid management can have an impact on patient outcome. In a larger trial of 80 patients undergoing colorectal surgery, MacKay et al.45 did not confirm these findings, despite their protocols for postoperative fluid management seeming, at first, comparable. However, a decision analysis reveals any comparison between these two studies to be difficult: Patients of both randomization groups were intraoperatively treated with relative fluid restriction (basal rate 10 ml \cdot kg⁻¹ \cdot h⁻¹) compared with the work of Lobo et al.⁴⁴ (approximately 18 ml \cdot kg⁻¹ \cdot h⁻¹). This is clearly reflected by the postoperative weight gain, an indirect measure for the interstitial fluid shift. The patients in the restrictive group of MacKay et al. had a body weight decrease of 0.5 kg, whereas the increase in their liberally treated group (+0.7 kg) was even less than that of the restrictive group of Lobo et al. (+1.1 kg). Above that, a sufficient postoperative fluid balance is not possible, because oral fluid intake was only "encouraged"45 and not reported by McKay et al. Nevertheless, with no patient receiving more than 3 l intravenous fluid a day, even perioperatively, most likely even their standard group was treated too restrictively to cause measurable harm. These findings underline the importance of a rational concept comprising the entire perioperative treatment.

In a multicenter study, Brandstrup et al.⁸ investigated a homogenous collective of 141 patients undergoing major colorectal surgery. They demonstrated that perioperative intravenous fluid restriction (mean 2,740 vs. 5,388 ml) significantly reduced the incidence of major and minor complications, such as anastomotic leakage, pulmonary edema, pneumonia, and wound infection. Despite limited fluid application and a perioperative decrease in urine output, acute renal failure did not occur in any patient. However, Brandstrup et al. did not purely compare liberal versus restrictive, but, as a close look at the infusion protocols reveals, colloids versus crystalloids, applying mainly colloids to the restrictive group while treating the liberal group with more than 5 l crystalloids.⁸ Nisanevic et al. found decreased postoperative morbidity, including a shortened hospital stay, under a protocol-based, more restrictive fluid therapy (1.2 vs. 3.7 l) in a more heterogeneous collective consisting of 152 patients scheduled to undergo mixed abdominal surgery.11

As a conclusion of a systematic review of 80 randomized clinical trials, Holte and Kehlet⁴³ recently recommended avoiding "fluid overload in major surgical procedures."

On the other hand, there is also the point of view that liberal fluid therapy has beneficial effects on various outcome parameters. In the following passages, we examine these assertions more carefully.

Postoperative Nausea and Vomiting

Maharaj et al.,19 for example, reported large fluid amounts during laparoscopic surgery to decrease pain and PONV. However, their restrictively treated group received only 212 ml fluid perioperatively after a fasting period of 13 h. The patients with an increased incidence of PONV might not have received an adequate fluid replacement therapy to restore their extracellular compartment. Above that, 65% of the restrictively treated patients received morphine (vs. 35% in the liberally treated group) before hospital discharge, which itself is known to increase the risk of PONV.⁴⁶ Furthermore, this could mean that large fluid amounts have the potential to decrease postoperative pain, an important additional effect in this collective. Magner et al.47 found comparable beneficial effects on the incidence of PONV after a 19min laparoscopic intervention, having infused 1,900 ml within this short period of time. Holte et al.,48 however, relativized this aggressive approach by demonstrating such, at first view, beneficial effects after knee surgery to be related to a decreased coagulation state and postoperative weight gain, which was still existent 72 h after surgery. During laparoscopy, however, they found a liberal fluid handling (40 vs. 15 ml/kg) to decrease PONV and to improve postoperative lung function. An observed increased release of atrial natriuretic peptide,17 which might crucially influence the vascular barrier function (see section titled Perioperative Protection of the Endothelial Surface Layer), did not seem to be related to measurable harm after minor surgery. On the other hand, McCaul et al.49 showed that even a complete lack of any perioperative infusion did not increase the risk of PONV compared with infusing 1.1 l compound sodium lactate.

These data, despite being inconsistent, indicate that higher fluid amounts might reduce the risk of PONV and increase postoperative lung function after short operations. Nevertheless, most studies considered only one outcome parameter; therefore, the overall effect on the patient is hard to gauge, because other, potentially more serious parameters may be impacted adversely by the same treatment. These results seem interesting regarding certain collectives, e.g., outpatients during minor surgery, but they cannot account for larger surgery over several hours. Current evidence suggests that liberal fluid is a good idea where major trauma and fluid shifting are unlikely, but more careful fluid management may be beneficial in more stressful operations.

Wound Infection and Tissue Oxygenation

Wound infections are serious complications of surgery. Oxidative killing by neutrophils is the most important defense against pathogens causing surgical infections.⁵⁰ Because oxygen is the substrate for oxidative killing, the rate of bacterial killing depends on sufficient tissue oxygenation. Therefore, the risk of surgical wound infection is inversely related to tissue oxygenation,⁵¹ which is also an important substrate for tissue repair and wound healing,^{52,53} and influenced by various factors: Mild hypothermia triples the risk of infection by reducing tissue oxygenation, but conversely, supplemental perioperative oxygen halves the risk of infection by increasing tissue oxygenation.⁵⁴ However, even supplemental oxygen does not improve oxygenation in hypoperfused tissues.⁵¹ Therefore, adequate perfusion is required for rapid healing and optimal resistance to infection. Obviously, it is important to perioperatively maintain an adequate blood volume,¹³ being, in principle, defined as a blood volume enabling the circulation to sufficiently perfuse the tissues. In practice, adequate volume is usually defined by hemodynamic stability, because there is no routine clinical method for evaluating tissue perfusion. Because hypovolemia does not only reduce peripheral tissue perfusion before compromising blood pressure, increasing heart rate, or reducing urine output,⁵⁵ it is evident how important it is to avoid hypovolemia.

Several studies during major abdominal surgery postulate that "aggressive fluid administration" increases tissue oxygenation.^{53,55} Because potential danger resulting from crystalloid overload was not well studied until very recently, liberal fluid handling has been traditionally recommended by many textbooks. Unfortunately, the underlying studies have several shortcomings. Mostly, tissue oxygenation was the only reported outcome parameter; weight gain, edema formation, anastomosis healing, coagulation factors, hospital stay, bowel function, renal failure, and cardiopulmonary complications-all wellknown effects of excessive fluid overload^{8,11,44}-were not measured. Above that, in most studies, patients received bowel preparation the day before surgery, which is currently questioned,^{56,57} and fasted for more than 8 h, which also is not in accordance with current guidelines.^{58,59} Despite the fact that fasting alone only slightly decreases extracellular fluid but maintaining intravascular normovolemia,⁶⁰ a combination with preoperative bowel preparation is suspect to induce a significant intravascular deficit.^{9,10} Therefore, most of the study patients were likely to be hypovolemic.⁶¹ Applying 2.1 versus 3.8 l crystalloids during major abdominal surgery (including the preoperative phase) after fasting and bowel preparation does not seem to be the intended conservative versus aggressive fluid therapy, but rather a too restrictive versus adequate regimen.53 What crystalloid overload could mean to the tissues was illustrated more than 30 yr ago by an animal experiment. Infusion of 10 ml/kg isotonic saline solution in rabbits significantly decreased tissue oxygen tension for 3.5 days.⁶² In a recent study from Kabon et al.,⁶³ patients receiving bowel preparation had substitution of 2 ml \cdot kg⁻¹ \cdot h⁻¹ overnight, and then 2.5 and 4.6 l crystalloid administration during major abdominal surgery were compared. No improvement of wound infection or wound healing was found after colorectal surgery in the "aggressively" treated group. Recently, Kimberger et al.64 showed that tissue oxygen tension can be increased by supplemental oxygen but not by supplemental crystalloid fluid, whereas Hiltebrand et al.65 found no augmentation in tissue oxygen pressure by high versus medium or low fluid regimens. While these are experimental data from animals with various limitations, recent evidence suggests that aggressive fluid therapy can be detrimental even in humans. Nisanevich et al.11 reported a higher rate of infectious complications (including surgical site infection) and a longer hospitalization period for the group receiving a large volume of fluids.

Moreover, epidural anesthesia^{66,67} and mild hypercapnia^{68,69} have been shown to increase subcutaneous tissue oxygenation. Sufficient tissue perfusion has been shown to have a benefit on survival in high-risk patients and depend on a higher mean arterial pressure, cardiac index, and mixed venous oxygen saturation, as well as significantly higher oxygen delivery and oxygen consumption.⁷⁰ Intravascular volume replacement with colloids (hydroxyethyl starch 130/0.4) has been shown to reduce the inflammatory response in patients undergoing major surgery compared with a crystalloid-based volume therapy.¹⁴ This has been interpreted to be most likely due to an improved microcirculation with reduced endothelial activation and less endothelial damage.¹⁸

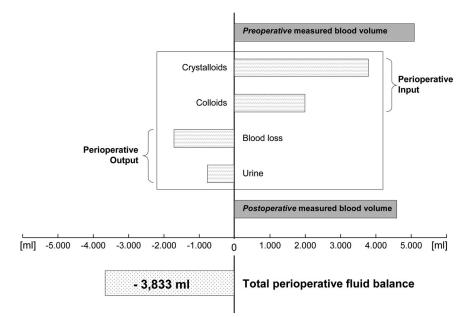
Because of a total lack of standardization, the available data do not allow evidence-based recommendations on practical perioperative fluid management.⁴³ Any perioperative fluid handling seems to be justified. However, this is in clear contrast to daily clinical observations during surgery, suggesting that our various surgical and anesthesiologic standard treatments might contribute to important perioperative problems.

Fluid Shifting: A Relevant Perioperative Problem

Fluid shifting out of the vasculature depends, in principle, on the body core temperature. Below 30°C, a significant decrease of plasma volume, accompanied by a decrease of central venous pressure, an increase of pulmonary and systemic resistance, and an increase of hematocrit have been reported.⁷¹ Between 37° and 33°C, however, no significant dependence on body temperature has been observed. Accordingly, this should not be a frequent intraoperative problem in noncardiac surgery. Nevertheless, fluid shifting is an often recognized phenomenon during and after surgical procedures. Direct and indirect blood volume measurements have shown that major surgery causes a deficit of 3-61 in the sensible perioperative fluid balance, 31, 32, 34, 72 *i.e.*, measurable input (crystalloid and colloid) minus measurable output (blood loss and urine output; fig. 1). This shift is not only an intraoperative but also a postoperative problem. The peak of fluid shifting has been reported to be at 5 h after trauma and to persist for up to 72 h, depending on the location of the operation site and on the duration of surgery.⁷³ Lowell et al.⁷⁴ found that 40% of patients admitted to a surgical intensive care unit had an excessive increase in body water of more than 10% from preoperative weight. Extracellular volume (ECV) overload has been shown to exceed 10 1 after 2 days of resuscitation in patients with sepsis. This fluid storage was obviously trapped inside the body and needed 3 weeks to be excreted.⁷⁵ Above that, even in healthy volunteers, it has been demonstrated that it takes 2 days to completely excrete a saline infusion of 22 ml/kg.^{76,77} One week after fluid resuscitation with 3-7 l fluid in patients with burns, only half of the patients had eliminated this infusion.⁷⁸ But not only the fluid shift out of the vasculature seems to be dangerous for patients. Also, fluid reabsorption can result in cardiac overload, occasionally leading to acute cardiac failure and pulmonary edema.79

Perioperative weight gain, being the most reliable marker of fluid storage outside the circulatory space, has been shown to be strongly related to patient mortality in a retrospective study of patients not randomly assigned to distinct fluid infusion regimens: In patients who gained less than 10% body weight, mortality was 10%; in patients whose body weight increased between 10% and 20%, mortality was 32%; and in patients whose body weight increased by more than 20%, mortality was 100% (fig. 2).⁷⁴ An unanswered question in this context is whether fluid shifting was a cause or an effect. However, the study impressively illustrated what was going on in the operating rooms of the late 1980s and what still frequently happens to patients in 2008. The operations investigated here were associated with severe trauma and blood loss. Nevertheless, the patients were treated

Fig. 1. Median blood volume status of 13 patients with ovarian cancer before and after major abdominal surgery, receiving a standard infusion regimen (crystalloids: approximately 12 ml \cdot kg⁻¹ \cdot h⁻¹; iso-oncotic colloids: blood loss replacement in ratio 1:1). Direct blood volume measurements (double-label technique) revealed a perioperatively occurring, at first unexplainable fluid loss out of the circulation.97 Median values (range): Preoperative blood volume 5,104 (4,099-6,004) ml, crystalloid infusion 3,800 (800-8,000) ml. colloid infusion 2.000 (0-4.700) ml. blood loss 1,700 (100-3,800) ml, urine production 750 (100-1,950) ml, postoperative blood volume 4,621 (3,802-5,170) ml. Total perioperative fluid balance is calculated from the measured parameters.



with large amounts of crystalloids and-if necessaryblood products. This means that a loss of colloid osmotic force was not sufficiently replaced. Patients with a relative low crystalloid infusion (4.1 l) had a weight gain of 4.7%, 1.7 days of postoperative ventilation, a vasopressor dependence of 2.8 days, and a mortality of 10%. In the group receiving "aggressive"⁷⁴ fluid resuscitation (12.51 crystalloids), weight gain accumulated to 31.7%, 6 days of postoperative ventilation were required, and 26 days of vasopressors therapy were necessary. Mortality was 100%. In this group, despite an excessive fluid supply, 33% developed acute renal failure (vs. 17% in the low fluid group). Also, invasive monitoring and postoperative ventilation correlated with the amount of infused fluids and the postoperative weight gain. Nevertheless, the authors stated that it was "almost certainly true that in many instances excessive volume was adminis-

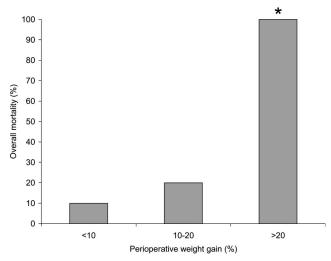


Fig. 2. Perioperative weight gain and mortality of patients. No patient survived if perioperative weight gain was more than 20%.⁷⁴ * *P* < 0.008 *versus* weight gain less than 10%.

tered."74 Other studies have shown a positive fluid balance in critically ill patients to be a common problem in the unit and to often be associated with poor outcome, such as increased mortality and prolonged intensive care treatment dependency and ventilator dependency.⁸⁰⁻⁸² Aggressive crystalloid infusion has been demonstrated to impede oxygen consumption, whereas lower infusion rates may provide better oxygen delivery with less increase of interstitial fluid accumulation.⁸³ A mean total applied amount of 12.5 l crystalloids, in combination with only 500 ml iso-oncotic colloid, but 18 units packed erythrocytes indicate a very hypooncotic therapy of an obviously excessive blood loss. Possibly, the patients would profit from a more colloid-accentuated resuscitation strategy, improving oxygen delivery and oxygen consumption by limiting extracellular fluid storage.⁸³

The corresponding "crystalloid versus colloid" debate has been enlarged by a "colloid versus colloid" controversy during the past years,⁸⁴ and one should obviously carefully distinguish what kind of colloid to use for which indication. Unfortunately, large randomized studies reliably comparing the two main colloids of interest, human albumin versus a modern, third-generation hydroxyethyl starch preparation, remain eagerly expected. The data regarding colloids versus crystalloid are contradictory. Hankeln et al.^{85,86} compared the cardiopulmonary effects of lactated Ringer's solution and 10% hydroxyethyl starch in 15 critically ill patients. Using the artificial colloid produced a significantly increased cardiac index, left and right ventricular stroke work index, central venous and wedge pressure, oxygen delivery, and oxygen consumption. Pulmonary vascular resistance index was reduced. The highly cited recent study by the investigators of Volume Substitution and Insulin Therapy in Severe Sepsis comparing 10% pentastarch (hydroxyethyl starch 200/0.5) with lactated Ringer's for resuscitation in

Anesthesiology, V 109, No 4, Oct 2008 Copyright By the American Society of Anesthesiologists. Unauthorized reproduction of this article is prohibited.

severe sepsis, by contrast, was stopped early because of a significant increase in acute renal failure in the group receiving hydroxyethyl starch.87 A careful look at the study design, however, reveals that, irrespective of whether bleeding occurred, a second-generation hypertonic hydroxyethyl starch at a total amount clearly beyond the recommended dose was applied. A subgroup analysis of low-dose ($\leq 22 \text{ ml/kg}$) versus high-dose (> 22 ml/kg) application revealed a significantly lower mortality (31% vs. 58%), even compared with crystalloid resuscitation (41%). Therefore, any conclusion drawn from these data regarding the use of hydroxyethyl starches in general should be made with caution. Nevertheless, these preparations are well known to induce severe side effects including pruritus, pleiotropic effects on the coagulation system, including reductions in coagulation factor levels, a decrease in number and function of platelets, and increased fibrinolysis.88 These effects, however, are clearly related to cumulative dose, mean molecular size, and substitution degree of the respective preparation.89,90

The use of human albumin versus crystalloid for fluid resuscitation did not improve outcome in 6,997 critically ill patients.^{91,92} In the subgroup of patients with traumatic brain injury, it actually increased mortality.93 Patients do not seem to inevitably benefit from colloid resuscitation if they do not experience bleeding or acute protein loss from the vasculature.⁹⁴

Generally, substantially more crystalloid is necessary to effect equivalent changes in hemodynamics. High-volume crystalloid resuscitation reduces oncotic pressure and may predispose to pulmonary⁹⁵ and peripheral edema, which interferes with tissue oxygen exchange and delays wound healing.96 However, not only crystalloids are shifted out of the vasculature, but also colloids (see section titled Crystalloid versus Colloid: Time to End an Erroneous Discussion).^{33,97}

This discussion raises several questions: Where does the body store this shifted fluid? Is it an interstitial shift or located within the mysterious third space? Does this space primarily consume fluid during major surgery which has to be replaced⁹⁸-or is fluid overload rather the trigger for such an occurrence? A small physiologic excursion into the mystic world surrounding the third space might provide some answers to these questions.

Interstitial or Third Space?

The third space has systematically been divided into an anatomical part and a nonanatomical part.99,100 Anatomical losses are considered to be a physiologic phenomenon at a pathologic amount, i.e., pathologic fluid accumulations within the interstitial space, the "functional" ECV (fECV). Physiologic fluid shifting with an intact vascular barrier from the vessels toward the interstitial space is considered to contain only small amounts of protein and only small molecules.¹⁰¹ As long as it is quantitatively managed by the lymphatic system, a physiologic shift does not cause interstitial edema.¹⁰² Overwhelming the lymphatic system, e.g., via excessive application of crystalloids, does. However, this problem can principally be resolved contemporarily via redistribution and urinary output.

Nonanatomical third space losses formally represent a fluid compartment functionally and anatomically separated from the interstitial space.^{98,103,104} Therefore, fluids trapped within this compartment are considered to now be part of the "nonfunctional" ECV (nfECV).¹⁰⁵ Losses toward this classic third space have been described as a fluid accumulation caused by major surgical procedures or trauma in spaces normally containing no or little fluid. Identified examples are the peritoneal cavity, the bowel, and traumatized tissues, but other, nonlocalized compartments are also postulated by the experts. Although total body water primarily remains unchanged by this theory, the "nonfunctional" part of extracellular fluid increases at the expense of the "functional" one. At least on the scene, this part is believed to be lost for extracellular exchange; it is unable to participate in the extracellular dynamic equilibrium.

Third Space: Quantification

Despite an intensive search for the perioperatively lost fluid, it is not localized in "nonanatomical" spaces: Neither the gut¹⁰⁶ nor traumatized tissue¹⁰⁷ contain these high amounts of fluid. Classic third space fluid losses have never been measured directly, and the actual location of the lost fluid remains unclear.¹⁰³ Therefore, these losses have been merely quantified indirectly by repeatedly measuring perioperative changes in the fECV via tracer-dilutional techniques, 103,108 presuming that the total ECV (functional plus nonfunctional) remains constant. These techniques are based on the principle of applying a known amount of a "suitable" tracer into a certain fluid compartment of the body. The concentration of the tracer within this compartment after a "suitable" equilibration interval leads to the distribution volume. The nfECV, however, is an ill-defined space, and high demands are made on a tracer to label fECV. On one hand, it must pass through the capillary wall membrane, but on the other hand, it must be excluded by the body's cell membranes, thus, making it extremely difficult to produce exact measurements. Such procedures, therefore, are limited by three main questions identifying three major shortcomings of tracer dilution when applied to fECV measurements: What is a suitable tracer distributing exclusively within the fECV? What is a suitable equilibration interval, allowing complete distribution, but not interfering with redistribution or tracer-

728

elimination kinetics? And, finally, how can a method to quantify fECV be reliably validated?

Despite these concerns, different tracers, techniques, sampling times, and mathematical calculations of the fECV have been used, leading to different results and various conclusions. It seemed that only trials using the sulfate tracer ${}^{35}SO_4$ with a relatively short equilibration time and calculating the fECV from a single or very few blood samples have reported a third space loss during surgery or hemorrhagic hypotension.¹⁰⁹⁻¹¹² Adequate equilibration times to measure fECV have, however, been reported to be up to 3 h for the sulfate¹¹³ and over 10 h for bromide (⁸²Br),¹¹⁴ the most common tracers. Problems of these tracers are that bromide enters erythrocytes and is excreted in bile,¹¹⁵ whereas sulfate is bound to plasma components¹¹⁶ and accumulates in the liver, in the kidneys, or during shock in muscular tissue.^{113,117} Above that, the necessary time for a tracer to achieve equilibration has been shown to be prolonged after surgery,^{104,118} hemorrhagic hypotension,^{104,118-120} and fluid overload.¹²¹ Single measurements will produce too-low or too-high plasma concentrations, overestimating or underestimating the calculated volume distribution if not taken at the exact equilibration time. Therefore, it is recommended to calculate tracer spaces from continued multiple samples until equilibration is shown in each individual case.^{108,122} Above that, a prerequisite for using tracer kinetics for volume measurements is a steady state condition, hardly given during shock or surgery.^{106,108,111,122} Surprisingly, trials using multiple blood samples after longer equilibration times to measure the fECV all found the opposite of a third space loss: After surgery, an unchanged or even increased fECV was detected.^{104,106,111,114,118,123-133} Trials using the bromide tracer all found an fECV expansion after surgery, unaccounted for by the calculated fluid balance.^{114,118,123,124,127,128} Accordingly, and in contrast to the common assumption, the majority of the data do not support the existence of a third space.

In summary, a classic third space was never localized and only "quantified" with one specific method using certain conditions regarding sampling and equilibration times, implying serious concerns and weaknesses.^{104,119,128,134} All other methods using various tracers, multiple sampling techniques, longer equilibration times, or analysis of kinetics contradict the existence of a fluid-consuming third space.^{104,114,118,123,124,128,129,133,135-139} Taking all this into account, we have to conclude that a classic third space per se quantitatively does not exist. It is currently not more than an ill-defined compartment thought to reflect an otherwise unexplainable perioperative fluid shift. Therefore, we suggest abolishing this mystery and sticking to the given facts: Fluid is perioperatively shifted within the functional extracellular compartment, from the intravascular toward the interstitial space.

Perioperative Fluid Shift: Trigger or Effect of Liberal Fluid Handling?

Currently, it seems unclear whether high infused amounts of fluid are the cause or the effect of an occurring shift toward the interstitium. In particular, it is still not known whether surgery and trauma cause the main part of an impressive primary fluid shift outward that must be treated with high amounts of fluid or whether, rather, an overwhelming infusion therapy causes severe perioperative problems that should be avoidable for the anesthesiologist. An interesting animal study performed more than 20 yr ago gave an important clue.¹⁰⁰ It was demonstrated in a rabbit model during enteral anastomosis that the surgical manipulation itself is enough to cause a significant increase of the interstitial water load by 5-10%, without any infusion therapy. An accompanying crystalloid infusion of 5 ml \cdot kg⁻¹ \cdot h⁻¹ doubled this edema. Surgery and trauma per se obviously have the power to cause a certain extent of fluid shifting, whereas crystalloid infusion therapy impressively influences its extent.

The following will further explore the interesting possibility that current liberal fluid therapy, in addition to surgery, is related to a significant perioperative imbalance of fluid homeostasis.

The Physiologic Background

Intracellular fluid comprises two thirds of the body water. The remaining one third, approximately 15 l in the normal adult, designates the ECV, consisting of the plasma (approximately 3 l), the interstitial space (approximately 12 l), and small amounts of so-called transcellular fluids, such as gastrointestinal secretion, cerebrospinal fluid, and ocular fluid.^{103,140} Because the latter fluid compartments are obviously, even under physiologic conditions, anatomically separated and not in a dynamic equilibrium with the other two parts, they are considered to be "nonfunctional." By contrast, the interstitial space and the plasma represent the "functional" extracellular space, 98,103 in which water and small solutes can easily exchange, a prerequisite for cell nutrition.

Fluid distribution within the human body is related to the distribution of osmotic active substances. The physiologic distribution is maintained by biologic barriers and oxygen-consuming ion pumps. The intact vascular barrier cannot be crossed by large molecules and proteins in relevant amounts.¹⁴¹ This is important because it enables the circulation to generate a positive intravascular blood pressure without unlimited fluid loss toward the interstitial space. Ernest Starling, a British physiologist, introduced his underlying classic model of the vascular barrier as early as 1896: Inside the vessels, the hydrostatic pressure is high, as is the colloid osmotic pressure.¹⁴² In contrast, according to this model, the interstitial space contains a low amount of proteins, whereas the hydrostatic pressure there is also low (fig. 3). The theoretical net result is a low filtration rate per unit of time, assembling to:

$$\mathbf{J}_{\mathbf{v}} = \mathbf{K}_{\mathbf{f}} ([\mathbf{P}_{\mathbf{c}} - \mathbf{P}_{\mathbf{i}}] - \sigma [\pi_{\mathbf{c}} - \pi_{\mathbf{i}}]),$$

where $J_v =$ net filtration; $K_f =$ filtration coefficient; $P_c =$ capillary hydrostatic pressure; P_i = interstitial hydrostatic pressure; σ = refection coefficient, π_c = capillary oncotic pressure; and π_i = interstitial oncotic pressure.

Accordingly, a sufficient plasma protein concentration should be necessary to provide a physiologically active inward directed force to successfully oppose the hydrostatic pressure gradient. Nevertheless, a small net fluid and protein shift out of the blood vessels occurs all the time, but is disposed in a timely manner from the interstitial space via the lymphatic system under physiologic conditions.143

According to this model, crystalloid overload should only cause a moderate fluid shift toward the interstitial space. The resulting increase in interstitial hydrostatic pressure, together with a dilution of the interstitially

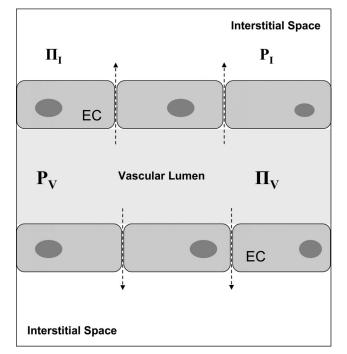


Fig. 3. The classic description of vascular barrier functioning in arterioles and capillaries, according to Ernest Starling (schematic): An inward-directed colloid-osmotic (= oncotic) pressure gradient is opposed to an outward-directed hydrostatic pressure of fluid and colloids. The arrows symbolize the small net fluid filtration assumed according to this model. The extremely simplified illustration does not consider the postulated small net fluid reabsorption on the venular site suggested by this model, due to an assumed decrease in the hydrostatic and an assumed increase in the oncotic pressure gradient. The Starling equation is mentioned in the main text. Π_{I} = oncotic pressure in the interstitial space; Π_v = oncotic pressure in the vascular lumen; EC = endothelial cell; P_v = hydrostatic pressure in the vascular lumen; $P_I =$ hydrostatic pressure in the interstitial space.

stored proteins, has been interpreted as an important edema-limiting mechanism.144 An accompanying increase in lymph flow in addition limits interstitial fluid volume expansion despite extracellular overload.¹⁰² Transfer of a substantial amount of the interstitial protein pool back into the vascular compartment by this increased lymph flow further contributes to this incident, increasing the inward-directed oncotic gradient.¹⁰² Surgery-induced inflammation, on the contrary, is believed to cause inevitable interruption and impediment of the reabsorption and return of the fluid to the circulation via the lymphatics.¹⁴⁵ This must derange the physiologic compensation and requires an intravascular fluid replacement by the anesthesiologist to maintain normovolemia and, therefore, cardiac preload. But does this way of thinking really reflect current physiologic knowledge? And is it really adequate to generalize fluids without distinguishing between crystalloids and colloids? A systematic look at perioperative losses and recent findings regarding the vascular barrier might help to answer this question.

Mechanics of Perioperative Fluid Handling

Perioperative fluid application basically must replace two kinds of losses: (1) losses occurring all the time (mainly urine production and insensible perspiration), possibly to another extent than under "normal" conditions; and (2) losses occurring exclusively during trauma and surgery (mainly blood losses). The first kind of loss affects the entire extracellular space, *i.e.*, the intravascular plus the interstitial space, and normally does not lead to a loss of colloid osmotic force from the intravascular space. The second loss induces a primarily isolated intravascular deficit, including losses of all blood components. In practice, we only have access to the vascular space, even when treatment of the entire extracellular compartment is intended.

Extracellular losses via urinary output and insensible perspiration are, schematically, replaced by absorption of colloid-free fluid and electrolytes from the gastrointestinal system. In the fasted patient, this compensation mechanism fails and has to be imitated artificially by the anesthesiologist. Theoretically, the best solution is an application of crystalloids, ideally in a balanced form, as not to cause acid-base disorders.¹⁴⁶ Because crystalloids are not retained at the vascular barrier after having been infused intravenously, they are homogenously distributed within the extracellular space, *i.e.*, four fifths are distributed into the interstitial space. Only one fifth remains intravascularly.

When substituting acute blood losses, there is no physiologic correlate we try to imitate, and each regimen must remain extemporaneous. Theoretically, an isovolemic online transfusion of warm whole blood should be considered

Anesthesiology, V 109, No 4, Oct 2008 opvriant © by the American Society of Anesthesiologists. Unauthorized reproduction of this article is prohibited.

optimal, because this is what is actually lost. Such an approach, however, is not an ideal target for perioperative treatment, because it has incalculable infectious and incompatibility risks.¹⁴⁷ Above that, it is expensive and suffers from insolvable logistical problems. Depending on the individual level of hemoglobin concentration, hemodilution improves blood rheology.¹⁴⁸ Therefore, hemodilution is not only a suitable alternative, but can be beneficial to the patient. Decreasing the hemoglobin value is, for example, what the circulation does during endurance training.¹⁴⁹ And this is the physiologic basis for preoperative normovolemic hemodilution to be applied to minimize the intraoperative transfusion rate.¹⁵⁰ Consequently, according to current knowledge, it is not inevitable to replace erythrocytes from the first milliliter of blood loss. Only a decrease below a certain hemoglobin value, individually depending on age, hemodynamic state, and previous illnesses, triggers a transfusion of concentrated erythrocytes.

Plasma components seem to be primarily dispensable as well. What might be correct for coagulation factors, however, has great impact when extended toward plasma proteins. According to the classic Starling concept, they must be maintained at a physiologic plasma concentration to preserve vascular barrier function.

Maintaining a physiologic state of the body fluid compartments as far as possible would mean a careful and adequate on-line substitution of actual fluid losses.

Interstitial Edema: The Price of Traditional Fluid Handling?

A conventional infusion regimen during major surgery is normally not based on physiologic facts but is predicated on, in doubt, liberal crystalloid handling. Application of artificial colloids is tolerated, but suspected to induce coagulation disorders, anaphylaxis, acute renal failure, and pruritus.^{87,151} Human albumin is currently not considered a suitable alternative for acute volume replacement in most countries, mainly for financial reasons. Accordingly, some textbooks still recommend coping with acute bleeding by an infusion of crystalloids at threefold to fourfold the actual blood loss.²⁸ An assumed hypovolemic state after fasting and a strong belief in an exorbitant insensible perspiration due to major surgery,13,152 together with a primarily fluid-consuming third space, leads to preoperative crystalloid loading (e.g., 2 ml/kg per hour of fasting).¹⁹ This is frequently followed by high basal crystalloid infusion rates of up to 15 ml \cdot kg⁻¹ \cdot h⁻¹ as a perioperative standard measure at least for major intraabdominal surgery.^{15,27} An increase in excretory kidney function from liberal crystalloid handling is expected and is an important argument to perform volume overload. From this point of view, fluid is only "offered" to the circulation and can easily be excreted if it falls into disuse. Also, a decreased circulatory state during

induction of general or neuraxial anesthesia is widely spread treated or even anticipated with fluid loading. Despite often being diagnosed as merely a relative hypovolemia due to a decrease of sympathetic tone, an infusion of crystalloids or colloids is considered to be harmless in contrast to vasopressor application, which threatens organ function, mainly that of the kidneys.^{153,154} Therefore, "clinicians are reluctant to use norepinephrine."154

The general aim to sufficiently substitute an assumed preoperative deficit and perioperative insensible losses (i.e., insensible perspiration plus fluid shifting out of the circulation) still leads to positive sensible fluid balances (i.e., blood loss and urinary output vs. infused fluids and blood products) of up to 101 at the end of major abdominal surgery.^{16,155-158} A related perioperative body weight gain at approximately the same extent^{16,155-159} indicates, however, that the contribution of the insensible perspiration to perioperative fluid needs should be small. And indeed, as early as in 1977, Lamke et al.¹⁵² performed direct measurements using a specially designed humidity chamber and clearly showed the insensible perspiration to be generally highly overestimated. The basal evaporation of approximately $0.5 \text{ ml} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$ in the awake adult increases to, at the most, 1 ml \cdot kg⁻¹ \cdot h⁻¹ during large abdominal surgery including maximal bowel exposure. Moreover, the impact of preoperative fasting on preoperative volume state is negligible: Even after an extended fasting period without concomitant bowel preparation, intravascular blood volume seems to be within normal ranges.⁶⁰ Anyway, the current fasting guidelines have more and more decreased the recommended preoperative period of no oral intake, at least for clear liquids,58,59 and bowel preparation is currently being severely questioned.56,57 Also, treatment of relative hypovolemia with volume instead of vasopressors, despite being in part successful if blood pressure is the only target,^{160,161} highly impacts the integrity of the body fluid compartments.

Intended volume expansion before induction of anesthesia via preoperative volume loading, despite still being widely performed, is at least questionable, because it accepts collateral damage: Crystalloids are physiologically distributed within the whole extracellular compartment, *i.e.*, as mentioned above, four fifths must leave the vasculature. This is illustrated by figure 4, indicating that the perioperatively infused amount of crystalloids corresponds to the perioperative weight gain. But also, isooncotic colloids do not completely remain within the circulatory compartment under such conditions as generally expected. Rather, to approximately 60% they do not expand blood volume but directly load the interstitial space.^{13,33} Infusing fluid not before but when relative hypovolemia occurs seems at first to be more reasonable, because volume effects of colloids have been demonstrated to be context sensitive.¹³ The volume effect is defined as that part of an infused bolus that does not shift outward but remains inside the vascula-

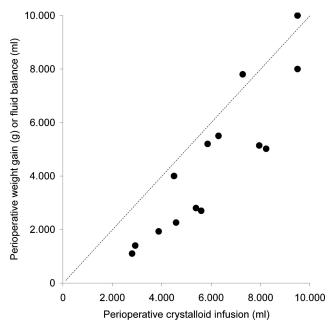


Fig. 4. Fifteen exemplary study groups suggest that perioperative weight gain increases with the perioperative amount of infused crystalloids.^{8,11,16,17,44,45,155–158,198} This illustrative diagram does not consider the number of patients in each study, only contains mean values without SD, and ignores intraoperative blood loss and insensible perspiration, as not reported in all studies. *Dashed line* = line of equality.

ture.³¹⁻³³ A simultaneous infusion of iso-oncotic colloids during acute bleeding, *i.e.*, when carefully maintaining intravascular normovolemia, led to volume effects of more than 90%.^{32,34} In contrast, approximately two thirds of an additional bolus of the same preparations in a normovolemic patient leaves the vasculature toward the interstitial space within minutes (fig. 5).³³ Consequently, volume effects of colloids depend on the "context," *i.e.*, the volume and hydration state of the patient.¹³ Above that, treating vasodilation with colloids ignores the fact that the cause of the intravascular volume expansion, an indirect vasodilatory effect of anesthetics, must be expected to terminate, *i.e.*, the vascular tone will be restored, at the end of surgery. Relative hypervolemia follows and occasionally causes postoperative pulmonary edema.¹⁴⁵ The kidney is not of much help in this situation: Because of significant surgical stress, the human body actively decreases excretory kidney function,^{9,162,163} obviously and reasonably to protect its fluid compartments.

Nevertheless, blood volume normally remains at preoperative levels even under generous fluid handling.^{31,32,164} This is in accord with a recently published mathematical description of perioperative fluid shifting during abdominal surgery. According to this model, intravenously applied crystalloids exceeding a certain level shift completely out of the circulation, loading the interstitial space.¹⁶⁴ This phenomenon is illustrated by the clinical observation that "prophylactic" crystalloid boluses in normovolemic patients have been shown to have no major effect on the incidence or severity of anesthesia-related hypotension in obstetric patients.^{165–168}

Consequently, interstitial edema is clearly the price for maintaining intravascular volume according to traditional recommendations, treating deficits that, in fact, do not exist with inadequate preparations. Colloids and crystalloids cannot be exchanged by simply adapting the amount. Actual losses from the vasculature are often treated by loading the entire extracellular compartment with crystalloids. But also the use of colloids does not always follow a rational concept.

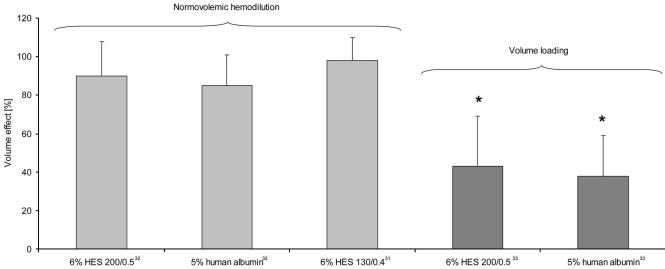


Fig. 5. The context sensitivity of volume effects of iso-oncotic colloids (the volume effect is that part of the colloid that remains within the circulation and does not primarily shift outward). As a substitute during acute bleeding, carefully maintaining normovolemia throughout the procedure, 6% hydroxyethyl starch (HES) 200/0.5, 5% human albumin, and 6% HES 130/0.4 (*left columns*) had volume effects of more than 90%. Volume loading of the normovolemic, by contrast, led to volume effects of 6% HES 200/0:5 and 5% human albumin (*right columns*) of approximately 40%. Blood volumes were assessed before and after intervention *via* double-label technique.^{31–33} n = 10 each. Values are mean ± SD. * P < 0.05 versus normovolemic hemodilution.

Downloaded From: https://anesthesiology.pubs.asahq.org/pdfaccess.ashx?url=/data/journals/jasa/931047/ on 04/14/2017

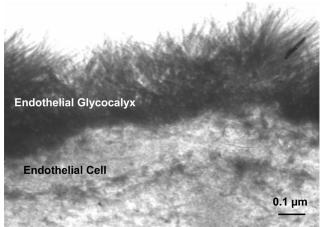


Fig. 6. Electron microscopic view of the endothelial glycocalyx. Staining of the glycocalyx was performed in modification of a method described by Vogel et al.,¹⁷⁷ based on an in situ stabilization of the glycocalyx by intracoronary application of a fixative containing lanthanum and glutaraldehyde.^{170,17}

The Shifts toward the Interstitial Space: The Beauty and the Beast

Fluid shifting toward the interstitial space can systematically be divided into two types: Type 1, the physiologic shift, occurs principally all the time. It represents an almost colloid-free shift of fluid and electrolytes out of the vasculature, occasionally at a pathologic amount, e.g., if large amounts of isotonic crystalloids are infused. This type of fluid shift occurs even if the vascular barrier is intact. Type 2, the pathologic shift, consists of fluids containing protein close to plasma concentration, crossing a functionally altered vascular barrier. The latter occurs inconstantly and is perioperatively related to the type, extent, and duration of surgery.¹⁰⁵ It is the result of two iatrogenic problems: first, a surgical one, increasing the protein permeability of capillaries and venules by up to eight times by endothelial damage due to mechanical stress, endotoxin exposure, ischemia-reperfusion injury, or inflammation¹⁶⁹; and second, an anesthesiologic one, having the power to in addition cause an impressive pathologic shift of protein and fluid toward the tissue in the context of acute hypervolemia.³³ The mechanism behind both phenomena seems to be an alteration of the endothelial glycocalyx.

The Endothelial Glycocalyx: The Gateway to the Interstitial Space

A healthy vascular endothelium is coated by the endothelial glycocalyx (fig. 6).¹⁷⁰⁻¹⁷³ This structure is a layer of membrane-bound proteoglycans and glycoproteins and was primarily regarded to have a thickness of only tens of nanometers.¹⁷⁴ Meanwhile, an endothelial surface

layer, consisting of the endothelial glycocalyx and bound plasma proteins and fluids, with a functional thickness of more than 1 μ m has been identified.^{173,175-177} This layer, together with the endothelial cells, is part of the doublebarrier concept of vascular permeability, identifying the glycocalyx as a second competent barrier in addition to the endothelial cell line opposing to unlimited extravasation.¹⁷³ By exerting a vital role on the physiologic endothelial permeability barrier^{176,178} and preventing leukocyte and platelet adhesion,179 it mitigates inflammation and tissue edema.¹⁷⁰⁻¹⁷²

The amount of plasma fixed within the endothelial surface layer and, therefore, quantitatively not participating in the normal blood circulation is approximately 700-1,000 ml in humans.^{32,172,180} However, this noncirculating part of plasma volume is in a dynamic equilibrium with the circulating part.¹⁷¹ Recently, it has been shown experimentally that a certain, fortunately small, minimal plasma concentration of albumin could represent a basic premise of the functional integrity of the endothelial surface layer.^{176,181}

The Starling Principle Meets the Endothelial Glycocalyx

The prerequisite for the classic Starling principle to be able to bind water within the vascular system is a significant colloid osmotic pressure gradient between the intravascular and extravascular space. However, several experiments have shown that this equation cannot be correct.182,183 The expected lymph flow, based on calculations according to the Starling principle, does not equal the measured flow.¹⁸⁴ Even after equilibration of intravascular and extravascular oncotic pressure in the isolated single microvessel model, the vascular barrier function remains intact.¹⁷⁸ There seems to be an oncotic gradient directly across the endothelial surface layer that defines vascular integrity, so that the presence of this layer should be the basic requirement for a physiologic barrier function.¹⁷⁶ In a rat mesenteric microvessel model, the effective colloid osmotic pressure difference opposing filtration was near 70% of the luminal osmotic pressure, though the colloid concentration outside equalled that inside the lumen of the microvessel.¹⁷⁸ It was proposed that the endothelial glycocalyx acts as a primary molecular filter and generates the effective oncotic gradient within a very small space.^{182,183,185} Transcapillary fluid exchange seems not to depend on the global difference between hydrostatic and oncotic pressure between blood and tissue. Rather, the hydrostatic and oncotic pressures between the blood and the small space directly underneath the endothelial glycocalyx, but still inside the anatomical lumen of the vessel, are decisive here (fig. 7).^{176,182,183} Taking the endothelial

surface layer into consideration, the Starling equation needs to be modified from its traditional form into:

$$J_{v} = K_{f}([P_{c} - P_{i}] - \sigma [\pi_{esl} - \pi_{b}]),$$

where $J_v =$ net filtration; $K_f =$ filtration coefficient; $P_c =$ capillary hydrostatic pressure; $P_i =$ interstitial hydrostatic pressure; σ = refection coefficient; π_{esl} = oncotic pressure within the endothelial surface layer; and $\pi_{\rm b}$ = oncotic pressure beneath the endothelial surface layer.

All this indicates a dependency between an alteration of the endothelial surface layer and protein or colloid shifting toward the interstitial space. Destruction of the endothelial surface layer and, therefore, the vascular barrier, leads back to the conditions proposed by the classic Starling equation, entailing transcapillary fluid shifting to equalize hydrostatic and oncotic pressures between tissue and blood-a catastrophe, if the interstitial colloid osmotic pressure equals that of the plasma.

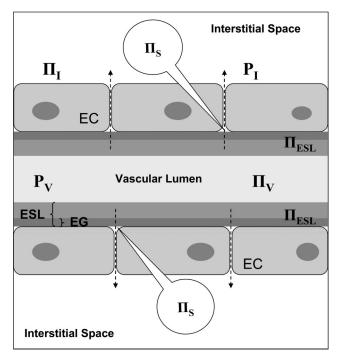


Fig. 7. The "revised" Starling principle.^{176,178} The hydrostatic pressure in the vascular lumen (Pv), which largely exceeds the interstitial pressure (P1), forces fluid outward. The endothelial glycocalyx (EG) binds plasma proteins, forming the endothelial surface layer (ESL) with a high internal oncotic pressure. The low net flux passing through the EG (arrows) has a sparse protein concentration; the oncotic pressure underneath the EG is low. Accordingly, an inward-directed oncotic pressure gradient develops just across the EG, while the proteins in the small space underneath the EG are continuously cleared toward the interstitial space via the remaining net flux. The extremely simplified illustration does not consider the venular site of the revised model, suggesting free and easy access of plasma pro-teins toward the interstitial space.¹⁷⁶ Because the hydrostatic force is low there, this should be no problem. Π_{ESL} = oncotic pressure within the endothelial surface layer; Π_{I} = oncotic pressure in the interstitial space; Π_s = oncotic pressure below the endothelial glycocalyx (subglyceal); Π_v = oncotic pressure in the vascular lumen; EC = endothelial cell.

This implies that, perioperatively, the endothelial glycocalyx should be preserved to inhibit a pathologic type 2 fluid shift into the interstitium. But how can this be achieved clinically?

Perioperative Protection of the Endothelial Surface Layer

Diminution of the endothelial glycocalyx leads to platelet aggregation,¹⁷⁹ leukocyte adhesion,¹⁸⁶ and an increase in endothelial permeability, causing tissue edema.^{170,176} According to experimental studies, ischemia-reperfusion,^{170,173} proteases,¹⁸⁷ tumor necrosis factor α ,¹⁸⁸ oxidized low-density lipoprotein,179 and atrial natriuretic peptide¹⁷⁵ have the power to degrade the endothelial glycocalyx. While surgical stress itself is well known to cause release of several inflammatory mediators,9,162,163 atrial natriuretic peptide release is triggered by iatrogenic acute hypervolemia.¹⁸⁹⁻¹⁹² This is in accord with the observation that intravascularly applied boluses of colloid increased the plasma protein filtration from the vascular bed in cardiopulmonary-healthy human subjects.³³ Obviously, despite not being easy to achieve, maintaining intravascular normovolemia could be the key in the hands of the anesthesiologist to protect the endothelial glycocalyx beyond the hardly avoidable damage caused by inflammatory mediators due to trauma and surgery. This could minimize pathologic fluid and protein shifts toward the interstitium via preservation of the endothelial glycocalyx. Above that, a certain minimal plasma protein content should be inevitable to form the endothelial surface layer in vivo.176,181

Crystalloid versus Colloid: Time to End an **Erroneous Discussion**

Recent comparisons of patient outcome after resuscitation using either crystalloids or colloids, irrespective of the actual reason of a decreased circulatory state,^{87,91,92} illustrate in an excellent manner what the so-called crystalloid versus colloid discussion¹⁻³ suffers from: Infusion solutions are generally not considered to be what they really are: drugs with indications, contraindications, and side effects. Ringer's solution as the only applied type of fluid during major abdominal surgery in humans decreased the mean tissue oxygen tension in the deltoid muscle for 24 h postoperatively by 23%, whereas additional treatment with hydroxyethyl starch led to a mean increase of 59%.¹⁸ Obviously, a considerable difference in interstitial architecture results from using colloids instead of crystalloids for volume replacement, and this seems quite logical. As extensively described above, isotonic crystalloids are distributed within the whole extracellular compartment, i.e., four fifths leave the vasculature, whereas iso-oncotic colloids have been designed to remain within the circulatory space. Consequently, the primary indication of crystalloids is replacement of fluid losses via (1) insensible perspiration and (2) urinary

output. Colloids, by contrast, are indicated to replace plasma deficits due to (2) acute blood loss or (2) proteinrich fluid shifts toward the interstitial space (pathologic type 2 shift).⁹⁷ Despite its being recommended²⁸ and still being widely performed, there is no rationale to substitute the first 1,000 ml of blood loss with a threefold to fourfold dose of isotonic crystalloids. Nor is there evidence to increase crystalloid infusion rate when patients seem to be clinically hypovolemic during surgery, despite intact extracellular fluid balance. This must induce an impressive (physiologic type 1) shift toward the interstitial space. Therefore, not only the amount, but also the kind, of applied fluid is crucial for patient outcome. Consequently, we must use the right kind of fluid in appropriate amounts at the right time to reduce collateral damage. From this point of view, it is erroneous to compare two classes of drugs with different indications regarding their impact on patient outcome.87,91,93,94 Rather, we must carefully distinguish between the different types of losses and treat them accordingly.

A discussion of this topic should, therefore, be focused on crystalloid and colloid-when to use what and at which amount, to minimize fluid shifting as far as possible.

Minimizing Type 1 Shifting (Crystalloid)

By using crystalloids as a substitute of acute blood losses, *i.e.*, infusing the entire extracellular space, interstitial edema is part of a questionable fluid concept and not a surprising accident. Accordingly, this type of fluid shifting should be minimized by using crystalloids only to replace urine production and insensible perspiration and by using iso-oncotic colloids for substitution of acute blood loss.

Minimizing Type 2 Shifting (Crystalloid and **Protein**)

To prevent this type of fluid shifting, it seems crucial to protect the endothelial surface layer. Perioperative alteration of this structure has two main causes: first, the release of inflammatory mediators due to surgical trauma; and second, the release of atrial natriuretic peptide during iatrogenic acute hypervolemia. Accordingly, some degree of interstitial edema seems to be unavoidable despite modification of perioperative fluid handling. The dimension of surgical stress-induced inflammation and vessel leakiness is proportional to the degree of injury.¹⁹³ The dimension of glycocalyx impairment has been shown to correspond to the surgical impact.¹⁹⁴ Even though a normal, well-controlled inflammatory response in a previously healthy patient almost always results in an uneventful recovery,¹⁹³ the resultant vessel leakiness seems to be inescapable. Nevertheless, an atraumatic surgical technique could be beneficial here.

Anesthesiologists might contribute to a reduction of stress release of inflammatory mediators by using neuraxial blocks. The endocrine response to surgery consists of an increased secretion of catabolically active hormones, most importantly cortisol, glucagon, and catecholamines.^{195,196} Single-dose neural blockade, applied as either intraoperative epidural or spinal anesthesia, has only a transient stress-reducing effect, without prolonged endocrine or metabolic effects.¹⁹⁶ Above that, epidural blockade is only partially effective in blocking the endocrine-metabolic responses after upper body procedures, because not everything is affected by epidural blockade.^{195,197} Single-shot blocks such as spinal anesthesia cannot achieve a sufficient reduction of inmediators.¹⁹⁶ Therefore, flammatory continuous neuraxial analgesia over 48-72 h using local anesthetics seems to be a possibility to reduce the metabolic stress response.

Nevertheless, carefully maintaining intravascular volume without hypervolemic peaks as far as possible currently seems to be the most promising concept. Prophylactic fluid boluses to anticipate acute bleeding or to extend intravascular blood volume in a primary normovolemic patient should no longer be considered state-of-the-art.

The situation changes when the glycocalyx is deteriorated during inflammation, ischemia, sepsis, or hypervolemia. Theoretically, colloids are only partly reflected at the vascular barrier in this situation. Despite the fact that there are no scientific data supporting this, many anesthesiologists use crystalloids in this situation. But is it really justified to consider an intentional load of the interstitial space with crystalloids to be the best strategy? Or is it rather inadequate? The shift out of a leaky vasculature is protein rich, and causal treatment of the intravascular deficit means an infusion of colloid osmotic force. Despite that this would lead to a shift quantitatively comparable with that when infusing crystalloid if the vascular barrier were completely open (which should be a rare case), it should be tried. Achieving a restoration of the circulating blood volume by infusing colloids to maintain intravascular normovolemia would reduce the interstitial load even if there is only a rudimentary competence of the vascular barrier. A central question in this context is: Which colloid is inert when it enters the interstitial space at a high amount, and which is not? The interstitial albumin concentration, for example, does not seem to differ relevantly from that which can be found in plasma, even under normal conditions.¹⁸³ Therefore, this natural colloid is an option in septic patients. However, future scientific efforts should concentrate on the less expensive, artificial alternatives.

During major surgery, it is nearly impossible to maintain normovolemia without producing edema. However, the occurring intravascular hypovolemia due to a protein-rich type 2 shift toward the tissue should be treated causally, i.e., with colloids. Using crystalloids in this situation aggravates this pathologic by a physiologic type 1 shift, which further increases the interstitial load. There are no clinical data supporting such an approach and, from a theoretical standpoint, it might even worsen the problem.

A Rational Approach to Perioperative Fluid Management

The goal of perioperative fluid application is the same than that of the cardiovascular system under normal conditions: an adequate blood flow in vital and, as far as possible, in traumatized tissues, as not to compromise the first and to enable effective wound healing in the latter. The focus of our efforts should be to avoid collateral damages, *i.e.*, interstitial edema, as far as possible. Therefore, it might be helpful to change our way of thinking from fluid "therapy" toward fluid "substitution." Above that, it is only a half-truth to proclaim a more restrictive therapy to be superior to a liberal one. Rather, an adequate and timely replacement of actual losses with appropriate preparations seems to be an ideal primary approach. Therefore, we should divide fluid therapy into two components: (1) replacement of fluid losses from the body via insensible perspiration and urinary output and (2) replacement of plasma losses from the circulation due to fluid shifting or acute bleeding. While a "goal-directed" approach via circulatory surrogates is, in principle, possible to replace plasma losses, the extracellular compartment cannot currently be monitored. Therefore, losses from the latter should be replaced based on a protocol:

- 1. The extracellular deficit after usual fasting is low.⁶⁰
- 2. The basal fluid loss *via* insensible perspiration is approximately 0.5 ml \cdot kg⁻¹ \cdot h⁻¹, extending to 1 ml \cdot kg⁻¹ \cdot h⁻¹ during major abdominal surgery.¹⁵²
- 3. A primarily fluid-consuming third space does not exist.

Plasma losses out of the circulation have to be replaced with iso-oncotic colloids, presuming the vascular barrier to be primarily intact and acknowledging that colloidal volume effects are context sensitive. The basis should be a timely replacement of visible blood losses, possibly supplemented by a goal-directed approach. Goals depend on local and individual circumstances and can vary from the maintenance of heart rate and blood pressure within a normal range in daily routine, up to stroke volume control via pulse pressure variation or esophageal Doppler in special cases. Importantly, despite being helpful, extended monitoring does not primarily seem to be the diagnostic hardware we urgently need to change in order to apply a more rational fluid concept. Rather, it seems warranted to replace the infusion of crystalloid by colloid if we detect the patient's circulation to be in need of additional volume.

Establishing a modern approach to perioperative fluid handling is currently hindered by the claim of successful studies to have treated their patients restrictively.^{8,11,44,45} Until recently, this has led to skepticism among clinicians, because many believe that restrictive fluid handling means depriving patients of their actual needs, leading to dehydration, which must, logically, lead to a decreased circulatory state due to intravascular hypovolemia. A careful comparison of the applied study protocols to measured values of preoperative blood volume after overnight fasting and insensible perspiration, however, reveals that the fluid regimens were mostly not restrictive in the true sense of the word, but represented an adequate substitution of fluid needs. A measurable weight gain even in restricted study groups^{8,11} indicates that there is still room for improvements in this context. To tap the full potential will be an important challenge in the next years.

Conclusion

We believe that a classic third space does not exist. Crystalloid overload, as well as iatrogenic deterioration of the vascular permeability barrier, can induce impressive fluid and protein shifting toward the interstitium. Consequently, and in accord with clinical studies, preoperative volume loading in normovolemic patients and routine replacement of high insensible and third space losses should be abolished in favor of demand-related fluid regimens. Fluid restriction in successful clinical outcome studies was not restrictive, but strongly related to the patient's actual losses. An adequate replacement of fluid needs seems to have the power to improve patient outcome and should be considered the therapy of choice to minimize perioperative fluid shifting.

The authors thank Bernhard F. Becker, M.D., Ph.D. (Professor of Physiology, Institute of Physiology, Ludwig-Maximilians University, Munich, Germany), for many years of close collaboration with experimental research on the endothelial glycocalyx and Ulrich Welsch, M.D., Ph.D. (Professor of Anatomy, Institute of Anatomy, Ludwig-Maximilians University), for providing electron microscopic pictures of the glycocalyx.

References

1. Bellomo R: Fluid resuscitation: Colloids versus crystalloids. Blood Purif 2002; 20:239-42

2. Choi PT, Yip G, Quinonez LG, Cook DJ: Crystalloids *versus* colloids in fluid resuscitation: A systematic review. Crit Care Med 1999; 27:200-10

3. Boldt J: Volume therapy in cardiac surgery: Are Americans different from Europeans? J Cardiothorac Vasc Anesth 2006; 20:98-105

4. Dorje P, Adhikary G, Tempe DK: Avoiding iatrogenic hyperchloremic acidosis: Call for a new crystalloid fluid. ANESTHESIOLOGY 2000; 92:625-6

5. Rehm M, Conzen PF, Peter K, Finsterer U: The Stewart model: "Modern" approach to the interpretation of the acid-base metabolism. Anaesthesist 2004; 53:347-57

6. Reid F, Lobo DN, Williams RN, Rowlands BJ, Allison SP: (Ab)normal saline and physiological Hartmann's solution: A randomized double-blind crossover study. Clin Sci (Lond) 2003; 104:17-24

7. Wakim KG: "Normal" 0.9 per cent salt solution is neither "normal" nor physiological. JAMA 1970; 214:1710

8. Brandstrup B, Tonnesen H, Beier-Holgersen R, Hjortso E, Ording H, Lindorff-Larsen K, Rasmussen MS, Lanng C, Wallin L, Iversen LH, Gramkow CS, Okholm M, Blemmer T, Svendsen PE, Rottensten HH, Thage B, Riis J, Jeppesen IS, Teilum D, Christensen AM, Graungaard B, Pott F: Effects of intravenous fluid restriction on postoperative complications: Comparison of two perioperative fluid regimens-A randomized assessor-blinded multicenter trial. Ann Surg 2003; 238: 641-8

9. Holte K, Sharrock NE, Kehlet H: Pathophysiology and clinical implications of perioperative fluid excess. Br J Anaesth 2002; 89:622-32

10. Holte K, Kehlet H: Compensatory fluid administration for preoperative dehydration: Does it improve outcome? Acta Anaesthesiol Scand 2002; 46: 1089-93

11. Nisanevich V, Felsenstein I, Almogy G, Weissman C, Einav S, Matot I: Effect of intraoperative fluid management on outcome after intraabdominal surgery. Anesthesiology 2005; 103:25-32

12. Wiedemann HP, Wheeler AP, Bernard GR, Thompson BT, Hayden D, deBoisblanc B, Connors AF Jr, Hite RD, Harabin AL: Comparison of two fluidmanagement strategies in acute lung injury. N Engl J Med 2006; 354:2564-75

13. Jacob M, Chappell D, Rehm M: Clinical update: Perioperative fluid management. Lancet 2007; 369:1984-6

14. Boldt J, Ducke M, Kumle B, Papsdorf M, Zurmeyer EL: Influence of different volume replacement strategies on inflammation and endothelial activation in the elderly undergoing major abdominal surgery. Intensive Care Med 2004; 30:416-22

15. Campbell IT, Baxter JN, Tweedie IE, Taylor GT, Keens SJ: IV fluids during surgery. Br J Anaesth 1990; 65:726-9

16. Dawidson IJ, Willms CD, Sandor ZF, Coorpender LL, Reisch JS, Fry WJ: Ringer's lactate with or without 3% dextran-60 as volume expanders during abdominal aortic surgery. Crit Care Med 1991; 19:36-42

17. Holte K, Klarskov B, Christensen DS, Lund C, Nielsen KG, Bie P, Kehlet H: Liberal versus restrictive fluid administration to improve recovery after laparoscopic cholecystectomy: A randomized, double-blind study. Ann Surg 2004; 240.892-9

18. Lang K, Boldt J, Suttner S, Haisch G: Colloids versus crystalloids and tissue oxygen tension in patients undergoing major abdominal surgery. Anesth Analg 2001; 93:405-9

19. Maharaj CH, Kallam SR, Malik A, Hassett P, Grady D, Laffey JG: Preoperative intravenous fluid therapy decreases postoperative nausea and pain in high risk patients. Anesth Analg 2005; 100:675-82

20. Waters JH, Gottlieb A, Schoenwald P, Popovich MJ, Sprung J, Nelson DR: Normal saline versus lactated Ringer's solution for intraoperative fluid management in patients undergoing abdominal aortic aneurysm repair: An outcome study. Anesth Analg 2001: 93:817-22

21. Farstad M, Haugen O, Rynning SE, Onarheim H, Husby P: Fluid shift is moderate and short-lived during acute crystalloid hemodilution and normothermic cardiopulmonary bypass in piglets. Acta Anaesthesiol Scand 2005; 49:949-55

22. Haugen O, Farstad M, Kvalheim V, Boe O, Husby P: Elevated flow rate during cardiopulmonary bypass is associated with fluid accumulation. J Thorac Cardiovasc Surg 2007; 134:587-93

23. Coe AJ, Revanas B: Is crystalloid preloading useful in spinal anaesthesia in the elderly? Anaesthesia 1990; 45:241-3

24. McCrae AF, Wildsmith JA: Prevention and treatment of hypotension during central neural block. Br J Anaesth 1993; 70:672-80

25. Nishimura N, Kajimoto Y, Kabe T, Sakamoto A: The effects of volume loading during epidural analgesia. Resuscitation 1985; 13:31-40

26. Pouta AM, Karinen J, Vuolteenaho OJ, Laatikainen TJ: Effect of intravenous fluid preload on vasoactive peptide secretion during Caesarean section under spinal anaesthesia. Anaesthesia 1996; 51:128-32

27. Sear JW: Kidney dysfunction in the postoperative period. Br J Anaesth 2005; 95:20-32

28. Kaye AD, Kucera AJ: Fluid and electrolyte physiology, Anesthesia, 6th edition. Edited by Miller RD. Philadelphia, Churchill Livingstone, 2005, pp 1763-98

29. Watenpaugh DE, Yancy CW, Buckey JC, Lane LD, Hargens AR, Blomqvist CG: Role of atrial natriuretic peptide in systemic responses to acute isotonic volume expansion. J Appl Physiol 1992; 73:1218-26

30. Noblett SE, Snowden CP, Shenton BK, Horgan AF: Randomized clinical trial assessing the effect of Doppler-optimized fluid management on outcome after elective colorectal resection. Br J Surg 2006; 93:1069-76

31. Jacob M, Rehm M, Orth V, Lotsch M, Brechtelsbauer H, Weninger E, Finsterer U: Exact measurement of the volume effect of 6% hydroxyethyl starch 130/0.4 (Voluven) during acute preoperative normovolemic hemodilution. Anaesthesist 2003; 52:896-904

32. Rehm M, Orth V, Kreimeier U, Thiel M, Haller M, Brechtelsbauer H, Finsterer U: Changes in intravascular volume during acute normovolemic hemodilution and intraoperative retransfusion in patients with radical hysterectomy. Anesthesiology 2000; 92:657-64

33. Rehm M, Haller M, Orth V, Kreimeier U, Jacob M, Dressel H, Mayer S, Brechtelsbauer H, Finsterer U: Changes in blood volume and hematocrit during acute preoperative volume loading with 5% albumin or 6% hetastarch solutions in patients before radical hysterectomy. ANESTHESIOLOGY 2001; 95:849-56

34. Rehm M, Orth VH, Kreimeier U, Thiel M, Mayer S, Brechtelsbauer H, Finsterer U: Changes in blood volume during acute normovolemic hemodilution with 5% albumin or 6% hydroxyethylstarch and intraoperative retransfusion. Anaesthesist 2001; 50:569-79

Downloaded From: https://anesthesiology.pubs.asahq.org/pdfaccess.ashx?url=/data/journals/jasa/931047/ on 04/14/2017

35. Goy RW, Chiu JW, Loo CC: Pulse dye densitometry: A novel bedside monitor of circulating blood volume. Ann Acad Med Singapore 2001; 30:192-8

36. Norberg A, Hahn RG, Li H, Olsson J, Prough DS, Borsheim E, Wolf S, Minton RK, Svensen CH: Population volume kinetics predicts retention of 0.9% saline infused in awake and isoflurane-anesthetized volunteers. ANESTHESIOLOGY 2007: 107:24-32

37. Sjostrand F, Hahn RG: Volume kinetics of glucose 2.5% solution during laparoscopic cholecystectomy. Br J Anaesth 2004; 92:485-92

38. Bendjelid K, Romand JA: Fluid responsiveness in mechanically ventilated patients: A review of indices used in intensive care. Intensive Care Med 2003; 29:352-60

39. Spahn DR, Chassot PG: Con: Fluid restriction for cardiac patients during major noncardiac surgery should be replaced by goal-directed intravascular fluid administration. Anesth Analg 2006; 102:344-6

40. Solus-Biguenet H, Fleyfel M, Tavernier B, Kipnis E, Onimus J, Robin E, Lebuffe G, Decoene C, Pruvot FR, Vallet B: Non-invasive prediction of fluid responsiveness during major hepatic surgery. Br J Anaesth 2006; 97:808-16

41. Tavernier B, Makhotine O, Lebuffe G, Dupont J, Scherpereel P: Systolic pressure variation as a guide to fluid therapy in patients with sepsis-induced hypotension. ANESTHESIOLOGY 1998; 89:1313-21

42. Gan TJ, Soppitt A, Maroof M, El-Moalem H, Robertson KM, Moretti E, Dwane P, Glass PS: Goal-directed intraoperative fluid administration reduces length of hospital stay after major surgery. ANESTHESIOLOGY 2002; 97:820-6

43. Holte K, Kehlet H: Fluid therapy and surgical outcomes in elective surgery: A need for reassessment in fast-track surgery. J Am Coll Surg 2006; 202:971-89

44. Lobo DN, Bostock KA, Neal KR, Perkins AC, Rowlands BJ, Allison SP: Effect of salt and water balance on recovery of gastrointestinal function after elective colonic resection: A randomised controlled trial. Lancet 2002; 359: 1812-8

45. MacKay G, Fearon K, McConnachie A, Serpell MG, Molloy RG, O'Dwyer PJ: Randomized clinical trial of the effect of postoperative intravenous fluid restriction on recovery after elective colorectal surgery. Br J Surg 2006; 93: 1469 - 74

46. Gan TJ: Risk factors for postoperative nausea and vomiting. Anesth Analg 2006; 102:1884-98

47. Magner JJ, McCaul C, Carton E, Gardiner J, Buggy D: Effect of intraoperative intravenous crystalloid infusion on postoperative nausea and vomiting after gynaecological laparoscopy: Comparison of 30 and 10 ml kg(-1). Br J Anaesth 2004; 93:381-5

48. Holte K, Kristensen BB, Valentiner L, Foss NB, Husted H, Kehlet H: Liberal versus restrictive fluid management in knee arthroplasty: A randomized, doubleblind study. Anesth Analg 2007; 105:465-74

49. McCaul C, Moran C, O'Cronin D, Naughton F, Geary M, Carton E, Gardiner J: Intravenous fluid loading with or without supplementary dextrose does not prevent nausea, vomiting and pain after laparoscopy. Can J Anaesth 2003; 50:440-4

50. Babior BM: Oxygen-dependent microbial killing by phagocytes (first of two parts). N Engl J Med 1978; 298:659-68

51. Gottrup F, Firmin R, Rabkin J, Halliday BJ, Hunt TK: Directly measured tissue oxygen tension and arterial oxygen tension assess tissue perfusion. Crit Care Med 1987; 15:1030-6

52. Prockop DJ, Kivirikko KJ, Tuderman L, Guzman NA: The biosynthesis of collagen and its disorders (first of two parts). N Engl J Med 1979; 301:13-23

53. Arkilic CF, Taguchi A, Sharma N, Ratnaraj J, Sessler DI, Read TE, Fleshman JW, Kurz A: Supplemental perioperative fluid administration increases tissue oxygen pressure. Surgery 2003; 133:49-55

54. Greif R, Akca O, Horn EP, Kurz A, Sessler DI: Supplemental perioperative oxygen to reduce the incidence of surgical-wound infection. Outcomes Research Group. N Engl J Med 2000; 342:161-7

55. Jonsson K, Jensen JA, Goodson WH III, West JM, Hunt TK: Assessment of perfusion in postoperative patients using tissue oxygen measurements. Br J Surg 1987: 74:263-7

56. Mariette C, Alves A, Benoist S, Bretagnol F, Mabrut JY, Slim K: Perioperative care in digestive surgery: Guidelines for the French Society of Digestive Surgery (SFCD). Ann Chir 2005; 130:108-24

57. Contant CM, Hop WC, van't Sant HP, Oostvogel HJ, Smeets HJ, Stassen LP, Neijenhuis PA, Idenburg FJ, Dijkhuis CM, Heres P, van Tets WF, Gerritsen JJ, Weidema WF: Mechanical bowel preparation for elective colorectal surgery: A multicentre randomised trial. Lancet 2007: 370:2112-7

58. Practice guidelines for preoperative fasting and the use of pharmacologic agents to reduce the risk of pulmonary aspiration: Application to healthy patients undergoing elective procedures-A report by the American Society of Anesthesiologist Task Force on Preoperative Fasting. ANESTHESIOLOGY 1999; 90:896-905

59. Soreide E, Eriksson LI, Hirlekar G, Eriksson H, Henneberg SW, Sandin R, Raeder J: Pre-operative fasting guidelines: An update. Acta Anaesthesiol Scand 2005; 49:1041-7

60. Jacob M, Chappell D, Conzen P, Finsterer U, Rehm M: Blood volume is normal after preoperative overnight fasting. Acta Anaesthesiol Scand 2008; 52: 522-9

61. Junghans T, Neuss H, Strohauer M, Raue W, Haase O, Schink T, Schwenk W: Hypovolemia after traditional preoperative care in patients undergoing colonic surgery is underrepresented in conventional hemodynamic monitoring. Int J Colorectal Dis 2006; 21:693-7

62. Heughan C, Ninikoski J, Hunt TK: Effect of excessive infusion of saline solution on tissue oxygen transport. Surg Gynecol Obstet 1972; 135:257-60

63. Kabon B, Akca O, Taguchi A, Nagele A, Jebadurai R, Arkilic CF, Sharma N, Ahluwalia A, Galandiuk S, Fleshman J, Sessler DI, Kurz A: Supplemental intravenous crystalloid administration does not reduce the risk of surgical wound infection. Anesth Analg 2005; 101:1546-53

64. Kimberger O, Fleischmann E, Brandt S, Kugener A, Kabon B, Hiltebrand L, Krejci V, Kurz A: Supplemental oxygen, but not supplemental crystalloid fluid, increases tissue oxygen tension in healthy and anastomotic colon in pigs. Anesth Analg 2007; 105:773-9

65. Hiltebrand LB, Pestel G, Hager H, Ratnaraj J, Sigurdsson GH, Kurz A: Perioperative fluid management: Comparison of high, medium and low fluid volume on tissue oxygen pressure in the small bowel and colon. Eur J Anaesthesiol 2007; 24:927-33

66. Buggy DJ, Doherty WL, Hart EM, Pallett EJ: Postoperative wound oxygen tension with epidural or intravenous analgesia: A prospective, randomized, single-blind clinical trial. ANESTHESIOLOGY 2002; 97:952-8

67. Treschan TA, Taguchi A, Ali SZ, Sharma N, Kabon B, Sessler DI, Kurz A; The effects of epidural and general anesthesia on tissue oxygenation. Anesth Analg 2003; 96:1553-7

68. Akca O, Doufas AG, Morioka N, Iscoe S, Fisher J, Sessler DI: Hypercapnia improves tissue oxygenation. ANESTHESIOLOGY 2002; 97:801-6

69. Hager H, Reddy D, Mandadi G, Pulley D, Eagon JC, Sessler DI, Kurz A: Hypercapnia improves tissue oxygenation in morbidly obese surgical patients. Anesth Analg 2006; 103:677-81

70. Shoemaker WC, Thangathurai D, Wo CC, Kuchta K, Canas M, Sullivan MJ, Farlo J, Roffey P, Zellman V, Katz RL: Intraoperative evaluation of tissue perfusion in high-risk patients by invasive and noninvasive hemodynamic monitoring. Crit Care Med 1999; 27:2147-52

71. Hammersborg SM, Farstad M, Haugen O, Kvalheim V, Onarheim H, Husby P: Time course variations of haemodynamics, plasma volume and microvascular fluid exchange following surface cooling: An experimental approach to accidental hypothermia. Resuscitation 2005; 65:211-9

72. Perko MJ, Jarnvig IL, Hojgaard-Rasmussen N, Eliasen K, Arendrup H: Electric impedance for evaluation of body fluid balance in cardiac surgical patients. J Cardiothorac Vasc Anesth 2001; 15:44-8

73. Robarts WM: Nature of the disturbance in the body fluid compartments during and after surgical operations. Br J Surg 1979; 66:691-5

74. Lowell JA, Schifferdecker C, Driscoll DF, Benotti PN, Bistrian BR: Postoperative fluid overload: Not a benign problem. Crit Care Med 1990; 18:728-33

75. Cheng AT, Plank LD, Hill GL: Prolonged overexpansion of extracellular water in elderly patients with sepsis. Arch Surg 1998; 133:745-51

76. Drummer C, Heer M, Baisch F, Blomqvist CG, Lang RE, Maass H, Gerzer R: Diuresis and natriuresis following isotonic saline infusion in healthy young volunteers before, during, and after HDT. Acta Physiol Scand Suppl 1992; 604: 101-11

77. Drummer C, Gerzer R, Heer M, Molz B, Bie P, Schlossberger M, Stadaeger C, Rocker L, Strollo F, Heyduck B: Effects of an acute saline infusion on fluid and electrolyte metabolism in humans. Am J Physiol 1992; 262:F744-54

78. Gump FE, Kinney JM, Iles M, Long CC: Duration and significance of large fluid loads administered for circulatory support. J Trauma 1970; 10:431-9

79. Salomon F: Acute dyspnea in fluid overload: Pathogenesis and differential diagnosis [in German]. Schweiz Med Wochenschr 1994; 124:1173-6

80. Humphrey H, Hall J, Sznajder I, Silverstein M, Wood L: Improved survival in ARDS patients associated with a reduction in pulmonary capillary wedge pressure. Chest 1990; 97:1176-80

81. Schuller D, Mitchell JP, Calandrino FS, Schuster DP: Fluid balance during pulmonary edema: Is fluid gain a marker or a cause of poor outcome? Chest 1991; 100:1068-75

82. Simmons RS, Berdine GG, Seidenfeld JJ, Prihoda TJ, Harris GD, Smith JD, Gilbert TI, Mota E, Johanson WG Jr: Fluid balance and the adult respiratory distress syndrome. Am Rev Respir Dis 1987; 135:924-9

83. Hauser CJ, Shoemaker WC, Turpin I, Goldberg SJ: Oxygen transport responses to colloids and crystalloids in critically ill surgical patients. Surg Gynecol Obstet 1980; 150:811-6

84. Boldt J, Scholhorn T, Mayer J, Piper S, Suttner S: The value of an albuminbased intravascular volume replacement strategy in elderly patients undergoing major abdominal surgery. Anesth Analg 2006; 103:191-9

85. Hankeln K, Siebert-Spelmeyer C, Bohmert F, Beez M, Laniewski P: Effect of colloid volume replacement substances and Ringer's lactate on hemodynamics and oxygen consumption of intensive care patients [in German]. Infusionstherapie 1988; 15:33-8

86. Hankeln K, Radel C, Beez M, Laniewski P, Bohmert F: Comparison of hvdroxvethyl starch and lactated Ringer's solution on hemodynamics and oxygen transport of critically ill patients in prospective crossover studies. Crit Care Med 1989: 17:133-5

87. Brunkhorst FM, Engel C, Bloos F, Meier-Hellmann A, Ragaller M, Weiler N, Moerer O, Gruendling M, Oppert M, Grond S, Olthoff D, Jaschinski U, John S, Rossaint R, Welte T, Schaefer M, Kern P, Kuhnt E, Kiehntopf M, Hartog C, Natanson C, Loeffler M, Reinhart K: Intensive insulin therapy and pentastarch resuscitation in severe sepsis. N Engl J Med 2008; 358:125-39

88. de Jonge E, Levi M: Effects of different plasma substitutes on blood coagulation: A comparative review. Crit Care Med 2001; 29:1261-7

89. Boldt J, Haisch G, Suttner S, Kumle B, Schellhaass A: Effects of a new modified, balanced hydroxyethyl starch preparation (Hextend) on measures of coagulation. Br J Anaesth 2002; 89:722-8

90. Jungheinrich C, Neff TA: Pharmacokinetics of hydroxyethyl starch. Clin Pharmacokinet 2005; 44:681-99

91. Finfer S, Bellomo R, Boyce N, French J, Myburgh J, Norton R: A comparison of albumin and saline for fluid resuscitation in the intensive care unit. N Engl I Med 2004: 350:2247-56

92. Finfer S, Bellomo R, McEvoy S, Lo SK, Myburgh J, Neal B, Norton R: Effect of baseline serum albumin concentration on outcome of resuscitation with albumin or saline in patients in intensive care units: Analysis of data from the saline versus albumin fluid evaluation (SAFE) study. BMJ 2006; 333:1044

93. Myburgh J, Cooper J, Finfer S, Bellomo R, Norton R, Bishop N, Kai LS, Vallance S: Saline or albumin for fluid resuscitation in patients with traumatic brain injury. N Engl J Med 2007; 357:874-84

94. Jacob M, Chappell D: Saline or albumin for fluid resuscitation in traumatic brain injury. N Engl J Med 2007; 357:2634-6

95. Rackow EC, Falk JL, Fein IA, Siegel JS, Packman MI, Haupt MT, Kaufman BS, Putnam D: Fluid resuscitation in circulatory shock: A comparison of the cardiorespiratory effects of albumin, hetastarch, and saline solutions in patients with hypovolemic and septic shock. Crit Care Med 1983: 11:839-50

96. Twigley AJ, Hillman KM: The end of the crystalloid era? A new approach to peri-operative fluid administration. Anaesthesia 1985; 40:860-71

97. Rehm M, Haller M, Brechtelsbauer H, Akbulut C, Finsterer U: Extra protein loss not caused by surgical bleeding in patients with ovarian cancer. Acta Anaesthesiol Scand 1998; 42:39-46

98. Shires T, Williams J, Brown F: Acute change in extracellular fluids associated with major surgical procedures. Ann Surg 1961; 154:803-10

99. Carrico CJ, Canizaro PC, Shires GT: Fluid resuscitation following injury: Rationale for the use of balanced salt solutions. Crit Care Med 1976; 4:46-54

100. Chan ST, Kapadia CR, Johnson AW, Radcliffe AG, Dudley HA: Extracellular fluid volume expansion and third space sequestration at the site of small bowel anastomoses. Br J Surg 1983; 70:36-9

101. Margarson MP, Soni N: Serum albumin: Touchstone or totem? Anaesthesia 1998; 53:789-803

102. Joles JA, Rabelink TJ, Braam B, Koomans HA: Plasma volume regulation: Defences against edema formation (with special emphasis on hypoproteinemia). Am J Nephrol 1993: 13:399-412

103. Brandstrup B: Fluid therapy for the surgical patient. Best Pract Res Clin Anaesthesiol 2006; 20:265-83

104. Roth E, Lax LC, Maloney JV Jr: Ringer's lactate solution and extracellular fluid volume in the surgical patient: A critical analysis. Ann Surg 1969; 169: 149-64

105. Woerlee GM: Common Perioperative Problems and the Anaesthetist. Dordrecht, Kluwer Academic Publishers, 1988

106. Nielsen OM: Extracellular fluid and colloid osmotic pressure in abdominal vascular surgery: A study of volume changes. Dan Med Bull 1991; 38:9-21

107. Doty DB, Hufnagel HV, Moseley RV: The distribution of body fluids following hemorrhage and resuscitation in combat casualties. Surg Gynecol Obstet 1970; 130:453-8

108. Brandstrup B. Svensen C. Engquist A: Hemorrhage and operation cause a contraction of the extracellular space needing replacement: Evidence and implications? A systematic review. Surgery 2006; 139:419-32

109. Carrico CJ, Coln CD, Lightfoot SA, Allsman A, Shires GT: Extracellular fluid volume replacement in hemorrhagic shock. Surg Forum 1963; 14:10-2

110. Shires T, Coln D, Carrico J, Lighfoot S: Fluid therapy in hemorrhagic shock. Arch Surg 1964; 88:688-93

111. Roberts JP, Roberts JD, Skinner C, Shires GT III, Illner H, Canizaro PC, Shires GT: Extracellular fluid deficit following operation and its correction with Ringer's lactate: A reassessment. Ann Surg 1985; 202:1-8

112. Fukuda Y, Fujita T, Shibuya J, Albert SN: The distribution between the intravascular and interstitial compartments of commonly utilized replacement fluids. Anesth Analg 1969; 48:831-8

113. Newton WT, Pease HD, Butcher HR Jr: Sodium and sulfate distributions in dogs after hemorrhagic shock. Surg Forum 1969; 20:1-2

114. Reid DJ: Intracellular and extracellular fluid volume during surgery. Br J Surg 1968: 55:594-6

115. Berson SA, Yalow RS: Critique of extracellular space measurements with small ions: Na24 and Br82 spaces. Science 1955; 121:34-6

116. Vineyard G, Osborne D: Simultaneous determination of extracellular water by 35-sulphate and 82-bromide in dogs, with a note on the acute effects of hypotensive shock. Surg Forum 1967; 18:37-9

117. Schloerb P, Peters C, Cage G, Kearns J, Lam J: Evaluation of the sulphate space as a measure of extracellular fluid. Surg Forum 1967; 18:39-41

118. Cleland J. Pluth JR. Tauxe WN. Kirklin JW: Blood volume and body fluid compartment changes soon after closed and open intracardiac surgery. J Thorac Cardiovasc Surg 1966; 52:698-705

119. Anderson RW, Simmons RL, Collins JA, Bredenberg CE, James PM, Levitsky S: Plasma volume and sulfate spaces in acute combat casualties. Surg Gynecol Obstet 1969; 128:719-24

120. Krejcie TC, Henthorn TK, Gentry WB, Niemann CU, Enders-Klein C, Shanks CA, Avram MJ: Modifications of blood volume alter the disposition of markers of blood volume, extracellular fluid, and total body water. J Pharmacol Exp Ther 1999; 291:1308-16

121. Herbst CA Jr: Simultaneous distribution rate and dilution volume of bromide-82 and thiocyanate in body fluid overload: Experimental and clinical correlation. Ann Surg 1974; 179:200-8

122. Jacob M. Conzen P. Finsterer U. Krafft A. Becker BF. Rehm M: Technical and physiological background of plasma volume measurement with indocyanine green: A clarification of misunderstandings. J Appl Physiol 2006; 102:1235-42

123. Breckenridge IM, Digerness SB, Kirklin JW: Validity of concept of increased extracellular fluid after open heart surgery. Surg Forum 1969; 20:169-71

124. Breckenridge IM, Digerness SB, Kirklin JW: Increased extracellular fluid after open intracardiac operation. Surg Gynecol Obstet 1970; 131:53-6

125. Gumpert JR, Zollinger RM, Riddell AG: Proceedings: The measurement of extracellular fluid volume with radiobromide simultaneous plasma and lymph disappearance in man. Br J Surg 1973; 60:903

126. Kragelund E: Loss of fluid and blood to the peritoneal cavity during abdominal surgery. Surgery 1971; 69:284-7

127. Pacifico AD, Digerness S, Kirklin JW: Acute alterations of body composition after open intracardiac operations. Circulation 1970; 41:331-41

128. Kragelund E: Changes of the apparent 3HOH, 82Br, 125I human albumin and 51Cr red blood cell dilution volumes before, during and after operation in human subjects. Ann Surg 1970; 172:116-24

129. Nielsen OM, Engell HC: Extracellular fluid volume and distribution in relation to changes in plasma colloid osmotic pressure after major surgery: A randomized study. Acta Chir Scand 1985; 151:221-5

130. Ladegaard-Pedersen HJ, Engell HC: A comparison of the distribution volumes of inulin and (51 Cr)EDTA in man and nephrectomized dogs. Scand J Clin Lab Invest 1972; 30:267-70

131. Gutelius JR, Shizgal HM, Lopez G: The effect of trauma on extracellular water volume. Arch Surg 1968; 97:206-14

132. Ladegaard-Pedersen HJ: Inulin distribution volume, plasma volume, and colloid osmotic pressure before and after major surgery. Acta Chir Scand 1974; 140:505-7

133. Shizgal HM, Solomon S, Gutelius JR: Body water distribution after operation. Surg Gynecol Obstet 1977; 144:35-41

134. Kragelund E, Dyrbye MO: Sulphate space in the human organism after intravenous administration of radiosulphate (Na2 358O4). Scand J Clin Lab Invest 1967.19.319-24

135. Albert SN, Shibuya J, Custeau P, Albert CA, Hirsch EF: A simplified method for measuring the volume of extracellular fluid by radioactive sulfur (\$35): Observations on shifts of fluid in induced hypotension. South Med J 1967; 60:933-9

136. Crystal RG, Baue AE: Influence of hemorrhagic hypotension on measurements of the extracellular fluid volume. Surg Gynecol Obstet 1969; 129:576-82

137. Furneaux RW, Tracy GD: The validity of the isotope dilution method of measuring extracellular fluid volume after acute haemorrhage. Aust J Exp Biol Med Sci 1970; 48:407-15

138. Moore FD, Dagher FJ, Boyden CM, Lee CJ, Lyons JH: Hemorrhage in normal man, I: Distribution and dispersal of saline infusions following acute blood loss-Clinical kinetics of blood volume support. Ann Surg 1966; 163:485-504

139. Shizgal HM, Lopez GA, Gutelius JR: Effects of experimental hemorrhagic shock on extracellular water volume. Ann Surg 1972; 176:736-41

140. Grocott MP, Mythen MG, Gan TJ: Perioperative fluid management and clinical outcomes in adults. Anesth Analg 2005; 100:1093-106

141. Stevens T, Garcia JG, Shasby DM, Bhattacharya J, Malik AB: Mechanisms regulating endothelial cell barrier function. Am J Physiol Lung Cell Mol Physiol 2000; 279:L419-22

142. Starling E: On the absorption of fluid from the connective tissue spaces. I Physiol (Lond) 1896; 19:312-26

143. Fleck A, Raines G, Hawker F, Trotter J, Wallace PI, Ledingham IM, Calman KC: Increased vascular permeability: A major cause of hypoalbuminaemia in disease and injury. Lancet 1985; 1:781-4

144. Lund T, Onarheim H, Reed RK: Pathogenesis of edema formation in burn injuries. World J Surg 1992; 16:2-9

145. Arieff AI: Fatal postoperative pulmonary edema: Pathogenesis and literature review. Chest 1999: 115:1371-7

146. Boldt J: The balanced concept of fluid resuscitation. Br J Anaesth 2007: 99:312-5

147. Klein HG, Spahn DR, Carson JL: Red blood cell transfusion in clinical practice. Lancet 2007; 370:415-26

148. Audibert G, Donner M, Lefevre JC, Stoltz JF, Laxenaire MC: Rheologic effects of plasma substitutes used for preoperative hemodilution. Anesth Analg 1994; 78:740-5

149. Brun JF, Bouchahda C, Chaze D, Benhaddad AA, Micallef JP, Mercier J: The paradox of hematocrit in exercise physiology: Which is the "normal" range from an hemorheologist's viewpoint? Clin Hemorheol Microcirc 2000; 22:287-303

150. Mielke LL, Entholzner EK, Kling M, Breinbauer BE, Burgkart R, Hargasser SR, Hipp RF: Preoperative acute hypervolemic hemodilution with hydroxyethylstarch: An alternative to acute normovolemic hemodilution? Anesth Analg 1997; 84:26-30

151. Kozek-Langenecker SA: Effects of hydroxyethyl starch solutions on hemostasis. Anesthesiology 2005; 103:654-60

152. Lamke LO, Nilsson GE, Reithner HL: Water loss by evaporation from the abdominal cavity during surgery. Acta Chir Scand 1977; 143:279-84

153. O'Brien EA, Bour SA, Marshall RL, Ahsan N, Yang HC: Effect of use of vasopressors in organ donors on immediate function of renal allografts. J Transpl Coord 1996; 6:215-6

154. Richer M, Robert S, Lebel M: Renal hemodynamics during norepinephrine and low-dose dopamine infusions in man. Crit Care Med 1996; 24:1150-6

155. Boldt J, Haisch G, Suttner S, Kumle B, Schellhase F: Are lactated Ringer's solution and normal saline solution equal with regard to coagulation? Anesth Analg 2002; 94:378-84

156. Shackford SR, Sise MJ, Fridlund PH, Rowley WR, Peters RM, Virgilio RW, Brimm JE: Hypertonic sodium lactate versus lactated ringer's solution for intravenous fluid therapy in operations on the abdominal aorta. Surgery 1983; 94: 41-51

157. Shackford SR, Fortlage DA, Peters RM, Hollingsworth-Fridlund P, Sise MJ: Serum osmolar and electrolyte changes associated with large infusions of hypertonic sodium lactate for intravascular volume expansion of patients undergoing aortic reconstruction. Surg Gynecol Obstet 1987; 164:127-36

158. Virgilio RW, Rice CL, Smith DE, James DR, Zarins CK, Hobelmann CF, Peters RM: Crystalloid versus colloid resuscitation: Is one better? A randomized clinical study. Surgery 1979; 85:129-39

159. Kudsk KA: Evidence for conservative fluid administration following elective surgery. Ann Surg 2003; 238:649-50

160. Ngan Kee WD, Khaw KS, Lee BB, Ng FF, Wong MM: Randomized controlled study of colloid preload before spinal anaesthesia for caesarean section. Br J Anaesth 2001; 87:772-4

161. Nishikawa K, Yokoyama N, Saito S, Goto F: Comparison of effects of rapid colloid loading before and after spinal anesthesia on maternal hemodynamics and neonatal outcomes in cesarean section. J Clin Monit Comput 2007; 21:125-9

162. Desborough JP: The stress response to trauma and surgery. Br J Anaesth 2000; 85:109-17

163. Wilmore DW: Metabolic response to severe surgical illness: Overview. World J Surg 2000; 24:705-11

164. Tatara T, Tashiro C: Quantitative analysis of fluid balance during abdominal surgery. Anesth Analg 2007; 104:347-54

165. Jackson R, Reid JA, Thorburn J: Volume preloading is not essential to prevent spinal-induced hypotension at caesarean section. Br J Anaesth 1995; 75:262-5

166. Karinen J, Rasanen J, Alahuhta S, Jouppila R, Jouppila P: Effect of crystalloid and colloid preloading on uteroplacental and maternal haemodynamic state during spinal anaesthesia for caesarean section. Br J Anaesth 1995; 75:531-5

167. Kinsella SM, Pirlet M, Mills MS, Tuckey JP, Thomas TA: Randomized study of intravenous fluid preload before epidural analgesia during labour. Br J Anaesth 2000; 85:311-3

168. Rout CC, Akoojee SS, Rocke DA, Gouws E: Rapid administration of crystalloid preload does not decrease the incidence of hypotension after spinal anaesthesia for elective caesarean section. Br J Anaesth 1992; 68:394-7

169. Landis EM: Heteroposity if the capillary wall as indicated by cinematographic analysis of the passage of dyes. Ann N Y Acad Sci 1964; 116:765-73

170. Chappell D, Jacob M, Hofmann-Kiefer K, Bruegger D, Rehm M, Conzen P, Welsch U, Becker BF: Hydrocortisone preserves the vascular barrier by protecting the endothelial glycocalyx. ANESTHESIOLOGY 2007; 107:776-84

171. Pries AR, Secomb TW, Gaehtgens P: The endothelial surface layer. Pflugers Arch 2000; 440:653-66

172. Pries AR, Kuebler WM: Normal endothelium. Handb Exp Pharmacol 2006; 1:1-40

173. Rehm M, Zahler S, Lotsch M, Welsch U, Conzen P, Jacob M, Becker BF: Endothelial glycocalyx as an additional barrier determining extravasation of 6% hydroxyethyl starch or 5% albumin solutions in the coronary vascular bed. ANESTHESIOLOGY 2004; 100:1211-23

174. Luft JH: Fine structures of capillary and endocapillary layer as revealed by ruthenium red. Fed Proc 1966; 25:1773-83

175. Bruegger D, Jacob M, Rehm M, Loetsch M, Welsch U, Conzen P, Becker BF: Atrial natriuretic peptide induces shedding of endothelial glycocalyx in coronary vascular bed of guinea pig hearts. Am J Physiol Heart Circ Physiol 2005; 289:H1993-9

176. Jacob M, Bruegger D, Rehm M, Stoeckelhuber M, Welsch U, Conzen P, Becker BF: The endothelial glycocalyx affords compatibility of Starling's principle and high cardiac interstitial albumin levels. Cardiovasc Res 2007; 73:575-86

177. Vogel J, Sperandio M, Pries AR, Linderkamp O, Gaehtgens P, Kuschinsky W: Influence of the endothelial glycocalyx on cerebral blood flow in mice. J Cereb Blood Flow Metab 2000; 20:1571-8

178. Adamson RH, Lenz JF, Zhang X, Adamson GN, Weinbaum S, Curry FE: Oncotic pressures opposing filtration across non-fenestrated rat microvessels. J Physiol 2004; 557:889-907

179. Vink H, Constantinescu AA, Spaan JA: Oxidized lipoproteins degrade the endothelial surface layer: Implications for platelet-endothelial cell adhesion. Circulation 2000; 101:1500-2

180. Nieuwdorp M, van Haeften TW, Gouverneur MC, Mooij HL, van Lieshout MH, Levi M, Meijers JC, Holleman F, Hoekstra JB, Vink H, Kastelein JJ, Stroes ES:

Loss of endothelial glycocalyx during acute hyperglycemia coincides with endothelial dysfunction and coagulation activation *in vivo*. Diabetes 2006; 55:480-6

181. Jacob M, Bruegger D, Rehm M, Welsch U, Conzen P, Becker BF: Contrasting effects of colloid and crystalloid resuscitation fluids on cardiac vascular permeability. ANESTHESIOLOGY 2006; 104:1223-31

182. Hu X, Weinbaum S: A new view of Starling's hypothesis at the microstructural level. Microvasc Res 1999; 58:281-304

183. Hu X, Adamson RH, Liu B, Curry FE, Weinbaum S: Starling forces that oppose filtration after tissue oncotic pressure is increased. Am J Physiol Heart Circ Physiol 2000; 279:H1724-36

184. Levick JR: Revision of the Starling principle: New views of tissue fluid balance. J Physiol 2004; 557:704

185. Michel CC: Starling: The formulation of his hypothesis of microvascular fluid exchange and its significance after 100 years. Exp Physiol 1997; 82:1-30

186. Constantinescu AA, Vink H, Spaan JA: Endothelial cell glycocalyx modulates immobilization of leukocytes at the endothelial surface. Arterioscler Thromb Vasc Biol 2003; 23:1541-7

187. Adamson RH: Permeability of frog mesenteric capillaries after partial pronase digestion of the endothelial glycocalyx. J Physiol 1990; 428:1-13

188. Henry CB, Duling BR: TNF-alpha increases entry of macromolecules into luminal endothelial cell glycocalyx. Am J Physiol Heart Circ Physiol 2000; 279: H2815-23

189. Kamp-Jensen M, Olesen KL, Bach V, Schutten HJ, Engquist A: Changes in serum electrolyte and atrial natriuretic peptide concentrations, acid-base and haemodynamic status after rapid infusion of isotonic saline and Ringer lactate solution in healthy volunteers. Br J Anaesth 1990; 64:606-10

190. Lewis H, Wilkins M, Selwyn B, Yelland U, Griffith M, Bhoola KD: Relationship between ANP, cyclic GMP and tissue kallikrein following saline infusion in healthy volunteers. Adv Exp Med Biol 1989; 247A:281-6

191. Schutten HJ, Johannessen AC, Torp-Pedersen C, Sander-Jensen K, Bie P, Warberg J: Central venous pressure: A physiological stimulus for secretion of atrial natriuretic peptide in humans? Acta Physiol Scand 1987; 131:265-72

192. Yamaji T, Ishibashi M, Takaku F: Atrial natriuretic factor in human blood. J Clin Invest 1985; 76:1705-9

193. Kohl BA, Deutschman CS: The inflammatory response to surgery and trauma. Curr Opin Crit Care 2006; 12:325-32

194. Rehm M, Bruegger D, Christ F, Thiel M, Conzen P, Jacob M, Chappell D, Stoeckelhuber M, Welsch U, Reichart B, Peter K, Becker BF: Shedding of the endothelial glycocalyx in patients undergoing major vascular surgery with global and regional ischemia. Circulation 2007; 116:1896-906

195. Ganapathy S, Murkin JM, Dobkowski W, Boyd D: Stress and inflammatory response after beating heart surgery *versus* conventional bypass surgery: The role of thoracic epidural anesthesia. Heart Surg Forum 2001; 4:323–7

 Holte K, Kehlet H: Epidural anaesthesia and analgesia: Effects on surgical stress responses and implications for postoperative nutrition. Clin Nutr 2002; 21:199-206

197. Segawa H, Mori K, Kasai K, Fukata J, Nakao K: The role of the phrenic nerves in stress response in upper abdominal surgery. Anesth Analg 1996; 82:1215-24

198. Marik PE, Iglesias J, Maini B: Gastric intramucosal pH changes after volume replacement with hydroxyethyl starch or crystalloid in patients undergoing elective abdominal aortic aneurysm repair. J Crit Care 1997; 12:51-5